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Eyerich, Kilian; Brown, Sara J.; Perez White, Bethany E.; Tanaka, Reiko J.; Bissonette, Robert; Dhar, Sandipan

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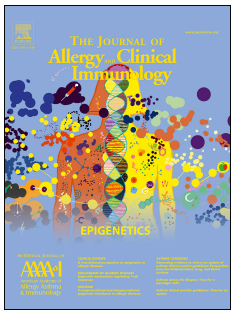

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Kilian Eyerich, MD, PhD, Sara J. Brown, MD, FRCPE, Bethany E. Perez White, PhD, Reiko J. Tanaka, PhD, Robert Bissonette, MD, Sandipan Dhar, MD, Thomas Bieber, MD, PhD, Dirk J. Hijnen, MD, PhD, Emma Guttman-Yassky, MD, PhD, Alan Irvine, MD, DSc, Jacob P. Thyssen, MD, PhD, DMSci, Christian Vestergaard, MD, PhD, DMSc, Thomas Werfel, MD, Andreas Wollenberg, MD, Amy S. Paller, MD, Nick J. Reynolds, BSc, MB BS, MD, FRCP

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Human and computational models of atopic dermatitis: a review and perspectives by an expert panel of the International Eczema Council

Kilian Eyerich, MD, PhD [1,2]#*, Sara J Brown, MD, FRCPE[3,4]#, Bethany E. Perez White, PhD[5]#, Reiko J. Tanaka, PhD[6]#, Robert Bissonette, MD[7], Sandipan Dhar, MD[8], Thomas Bieber, MD, PhD[9,10], Dirk J Hijnen, MD, PhD[11], Emma Guttman-Yassky, MD, PhD[12], Alan Irvine, MD, DSc[13], Jacob P. Thyssen, MD, PhD, DMSci[14], Christian Vestergaard, MD, PhD, DMSc[15], Thomas Werfel, MD[16], Andreas Wollenberg, MD[17], Amy S. Paller, MD[18]§, Nick J Reynolds BSc, MB BS, MD, FRCP[19,20]§*

#, §: these authors contributed equally to this work; *: corresponding authors

[1]Department of Dermatology and Allergy, Technical University of Munich, Germany; ²Center of Allergy and Environment (ZAUM), HMGU and Technical University of Munich, Germany; kilian.eyerich@tum.de; [3]Skin Research Group, School of Medicine, University of Dundee and [4]Department of Dermatology, Ninewells Hospital and Medical School, Dundee, United Kingdom; [5]Department of Dermatology and Skin Tissue Engineering Core, Feinberg School of Medicine, Northwestern University, Chicago, Illinois; [6]Department of Bioengineering, Imperial College London, London, United Kingdom; [7]Innovaderm Research Inc, Montreal, Quebec, Canada; [8]Department of Pediatric Dermatology, Institute of Child Health, Kolkata, West Bengal, India; [9]Department of Dermatology and Allergy, University of Bonn, Bonn, Germany; [10]Christine Kühne-Center for Allergy Research and Education, Davos, Switzerland; [11]Department of Dermatology, Erasmus University Medical Center (Erasmus MC), Rotterdam, The Netherlands; [12]Icahn School of Medicine at Mount Sinai Medical Center, New York, New York; [13]Trinity College Dublin, National Children's Research Centre, Paediatric Dermatology Our Lady's Children's Hospital, Dublin, Ireland, [14]Department of Dermatology and Allergy, National Allergy Research Centre, Herlev and Gentofte Hospital, University of Copenhagen, Denmark; [15]The Department of Dermatology, Aalborg Universityhospital, Aalborg

Denmark; [16] Medizinische Hochschule Hannover, Hannover, Germany; [17] Department of Dermatology and Allergy, Ludwig-Maximilian-University Munich, Munich, Germany; [18] Departments of Dermatology and Pediatrics and the Skin Disease Research Center, Northwestern University Feinberg School of Medicine, Chicago, Illinois; [19] Dermatological Sciences, Institute of Cellular Medicine, Newcastle University, Newcastle upon Tyne, UK; [20] Department of Dermatology, Royal Victoria Infirmary, Newcastle Hospitals NHS Foundation Trust, Newcastle upon Tyne, United Kingdom; nick.reynolds@ncl.ac.uk

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Abstract

Atopic dermatitis (AD) is a prevalent disease worldwide associated with systemic co-morbidities, representing a significant burden on individuals, their families and society. Therapeutic options for AD remain limited, in part due to lack of well-characterised animal models. To better define pathophysiological mechanisms and to identify novel therapeutic targets and biomarkers that predict therapeutic response, there has been increasing interest in developing experimental approaches to study the pathogenesis of human AD *in vivo*, *in vitro*, and *in silico*. This review critically appraises a range of models including: genetic mutations relevant to AD; experimental challenge of human skin *in vivo*; tissue culture models; integration of "omic" datasets; and the development of predictive computational models. Whilst no one individual model recapitulates the complex AD pathophysiology, our review highlights insights gained into key elements of cutaneous biology, molecular pathways and therapeutic target identification through each approach. Recent developments in computational analysis, including the application of machine learning and a systems approach to data integration and predictive modelling, highlight the applicability of these methods to AD subclassification (endotyping), therapy development and precision medicine. Such predictive modelling will highlight knowledge gaps, further inform refinement of biological models, and support new experimental and systems approaches to AD.

Key words: Atopic dermatitis, atopic eczema, Endotype**,** Human models, Machine learning**,** Mechanistic models**,** Precision medicine**,** Tissue culture models**,** Skin equivalents**,** Systems biology

Abbreviations

ACD	Allergic contact dermatitis
AD	Atopic dermatitis
APT	Atopy Patch Test
ILs	Interleukins
IRFs	Interferon regulatory factors

IPEX	Polyendocrinopathy Enteropathy X-linked syndrome
LV	Langerhans cells
PD	Pharmacodynamic
PK	Pharmacokinetic
RAST	Radioallergosorbent test
RNA-Seq	RNA-sequencing
SPT	Skin prick testing

Introduction

Atopic dermatitis (AD; synonym atopic eczema) has a complex aetiology, involving multiple genetic and environmental factors[1 2]. With its very high incidence in childhood, chronicity, devastating effect on quality of life for affected patients and their families, enormous socio-economic costs, and limited therapeutic options to date, AD represents a major challenge. Furthermore, there is clear evidence that AD represents a systemic inflammatory disease with multiple comorbidities extending beyond the well-recognized atopic associations[3]. Consequently, a number of animal models have been developed and utilized by investigators and the pharmaceutical industry to better understand the disease and consider new pathways to target[4]. However, as recently reviewed, mouse models do not adequately reflect the transcriptomic and gene pathways activated in human AD skin[5] and the intrinsic difference between mouse and human skin represents a barrier to direct translation of findings from animals into human disease. Consequently, there has been increasing interest in experimental studies in humans (in part facilitated by technological and "omic" developments), cell culture models utilizing human tissue, and the use of computational or mathematical models that are developed by integrating these data. In this review article, we have used the term "human AD model" to define representations of the disease state and interventions that enable scientific insight into disease pathogenesis, disease course, and response to therapy. We delineate and critically appraise these AD modelling approaches that range from the experimental study of human skin *in vivo* (including challenge studies and detailed phenotyping and investigation of patients harboring specific genetic mutations), the generation of AD-relevant models using immunological, genetic and molecular methods in 2D and 3D human tissue culture, to the development of *in silico* computational models using a systems biology approach. Whilst a reductionist approach cannot by definition recapitulate the full spectrum of AD, these models have greatly increased our understanding of the molecular drivers of AD and provide a powerful tool for preclinical drug development and target validation. However, just as the etiology, clinical expression, and severity of AD range broadly among patients, *in vitro* and *in silico* models of AD vary widely both in how the AD phenotype is induced and how the models are evaluated. Therefore, we invited members of the International Eczema Council (IEC; www.eczemacouncil.org), a group of experts

in AD, and associated authorities in the field to contribute to a scoping and development meeting and subsequently to evaluate and critically appraise the breadth of human AD and computational models to determine their strengths and weaknesses in how they recapitulate the pathophysiology of AD and enable therapeutics to be tested and validated.

***In vivo* models of AD**

To dissect the pathogenesis of AD, two general approaches using human *in vivo* models have been followed: i) the study of rare genetic variants with AD-like phenotypes; and ii) the experimental challenge of AD or non-AD subjects with allergens or irritants. Regarding the first approach, numerous studies have characterized genetic disorders that display skin barrier function abnormalities. Most often, these studies characterized ichthyosis vulgaris, a disease that allowed insights into the function of the epidermal differentiation gene *FLG* (encoding filaggrin), in which mutations show the strongest association to AD development of all known genes[6] (Figure 1). Other studies have focused on disorders characterized by systemic inflammation[3] and immunodeficiency with AD-like skin manifestations (Figure 1). One example is patients suffering from Immunodysregulation Polyendocrinopathy Enteropathy X-linked (IPEX) syndrome that serves as a model to study how systemic imbalances in the Treg population can drive cutaneous AD-like inflammation[7]. In addition, the link between type 2 immunity, transcription factors such as JAK or STAT, and high levels of IgE was investigated in immunodeficiency syndromes such as STAT3 and DOCK-8 hyper-IgE syndromes or combined immunodeficiency disorders[8, 9]. Table S1 lists the main genetic conditions that have provided insight into AD pathogenesis to date. Whilst the study of rare variants offers the opportunity to delineate distinct molecular mechanisms and control pathways of a particular phenotype, and thus may be regarded as “human models of AD”, a limitation of this approach is that not all observed phenomena are relevant in AD, which is more complex and heterogeneous than monogenic disorders.

The second *in vivo* approach to study the pathogenesis of AD is standardized challenge with allergens or other environmental factors. The most commonly used model is the Atopy Patch Test (APT), an

epicutaneous challenge of specific allergens dissolved in vehicle[10], which has provided insight into the temporal development of immune phenomena in AD[11] (Table S2). Although developed in part to define clinically relevant reactions to aero-allergens, food allergens and autoantigens[12 13 14], it´s validity and predictive value depend on a variety of factors in the protocol [15] and the APT is not used routinely in clinical practice. Experimentally, the APT has provided insights into the the temporal sequence of cutaneous cellular infiltrates. Acute skin lesions show a highly reproducible Th2 dominant inflitrate[16], although other cell types including Th17 cells are also present[17 18]. This Th2 dominance is in sharp contrast to other inflammatory skin diseases such as psoriasis[19, 20]. Time course studies have shown that additional immune cell subsets, such as Th1 and Th22 cells, progressively infiltrate the skin during an ongoing APT reaction, mirroring the cellular composition of acute versus chronic human AD[17 21]. The APT has also been used to characterize dendritic cells within early lesional AD skin, e.g. Inflammatory dendritic epidermal cells[18]. Furthermore, the APT has provided insights on the interaction of microbiota and our immune system, in particular the role of bacterial-derived superantigens acting as an amplifier of the allergen specific cutaneous response in AD[21, 22 23]. In all these experimental APT studies, the population of AD subjects were well defined with specific inclusion and exclusion criteria (although the precise definitions of AD varied); in most studies AD, together with specific IgE to the corresponding allergen used in the APT, was an inclusion criterion.

Hapten challenge to induce classical allergic contact dermatitis (ACD) in AD patients has also broadened our understanding of AD pathogenesis (Table S2). Whether AD patients have an increased risk of ACD remains controversial and may depend on whether they harbor *FLG* mutations, which may allowed increased penetration of allergens. However, attenuated ACD reactions have been reported in AD subjects compared to controls in a severity-dependent manner [24, 25]. This might be due to the fact that haptens induce distinct immune responses[26], with fragrances mimicking the Th2/Th22 dominance of AD while nickel, DNCB, or imiquimod[27] induced Th1/Th17 skewed immune responses. Of note, AD patients show a Th2-skewed ACD reaction[28], and this immune deviation might account for the diminished ACD prevalence in AD. A Th2

immune reaction profile of AD patients was also observed in an aero-challenge setup[29], as well as when challenging AD patients with physical factors such as hard water[30, 31].

All current challenge models have some limitations (Table S2), as they only represent acute reactions and the small areas of applications cannot reproduce the intense pruritus and sleep disturbances usually present in AD. Furthermore, to date they have not stratified for genetic differences/endotypes amongst AD patients comparing APTs in patients with and without *FLG* mutations, for example, might be a useful line of future investigation. Moreover, in the future, molecular profiling of lesional skin from standardized challenge models, adjusted according to AD endotype, might be used in early clinical studies to evaluate the potential of new drugs to improve AD[32].

In Vitro Models

As shown in Table S3, there are several 2D cell culture and 3D organotypic models for AD that complement each other in addressing specific experimental questions. While, 2D cell culture models (by definition) do not duplicate the architecture of skin, they are amenable to high-throughput techniques for drug discovery and target validation (2D model section, Supplementary Table S3). Accordingly, Otsuka *et al.* used 2D cultures to screen a chemical library for compounds that enhance *FLG* transcriptional activation and mRNA expression, suggesting a potential novel therapeutic agent for AD[33]. On the other hand, 3D models replicate the stratified, squamous epithelium of epidermis, but require specific expertise and are time consuming. Epidermal equivalents consist of keratinocytes without a dermal compartment, while skin equivalents have a dermis, such as fibroblast-embedded collagen (3D model section, Supplementary Table S3). Both 2D and 3D models are amenable to treatment with disease-relevant cytokines, gene knockdown, use of patient-derived cells, and/or co-culture (Figure 2 and Supplementary Table S3).

The immune system is a major driver of AD and *in vitro* immune modulation with disease-relevant cytokines, such as interleukins (ILs), can lead to AD-like phenotypes in normal primary keratinocytes[34] and

3D models [35-41] (3D cytokine model section, Supplementary Table S3). Knockdown of filaggrin in culture systems can give insight into the molecular and proteomic changes associated with its loss in AD[42]; and combining filaggrin knockdown with other perturbations, e.g., cytokine treatment, can be used to study the multifactorial drivers of AD. For example, Hönzke *et al.* reported that filaggrin knockdown exacerbated epidermal responses to IL-4 and 13, including increased proliferation and keratinocyte-released cytokines in 3D skin equivalents[43]. Patient-derived cells for 2D and 3D culture or tissue for explant culture are limited by access and availability, but may be the most relevant in terms of modeling AD [44-47]. Further, patient biopsies can be a source of skin cells other than keratinocytes, allowing for co-culture models. Given that multiple systems contribute to AD, co-culture models that include immune cells, dermal fibroblasts, and neurons can begin to address their interplay with keratinocytes. For example, Berroth *et al.* derived keratinocytes and fibroblasts from normal and AD skin and showed that AD-derived fibroblasts are sufficient to decrease *FLG* mRNA in normal-derived keratinocytes in 3D culture[47]. Moreover, combining *FLG* knockdown with CD4+ activated T-cells uncovered direct cross-talk between keratinocytes and T-cells that resulted in T-cell migration within the dermal compartment towards the epidermis [48]. These studies highlight the levels of complexity that can be engineered into the 3D culture models. 3D culture systems have also been used to understand environmental influences on skin, including air pollution, ultraviolet radiation exposure, and bacterial infection[49-51]. These relevant environmental factors could therefore be incorporated into *in vitro* models of AD. The 3D cultures and skin explants can also be used to assess the comparative efficacy and practical applicability of novel drug delivery systems [52, 53]. Notably, despite the assorted methodologies applied in developing *in vitro* models of AD, there is overlap in the AD-like characteristics amongst the various models: most produce perturbed epidermal morphology, abnormal differentiation, and barrier dysfunction. Most often, disparities in reported phenotypes appear to stem, at least in part, from differences in the methodologies used in evaluating models (not necessarily because of the absence of the phenotype).

Although *in vitro* models may not mimic certain symptomatic and/or subjective aspects of the disease such as pruritus and pain, they allow monitoring of changes in epidermal morphology and differentiation, gene and protein expression, lipid synthesis, and barrier function. Histologically, AD skin sections and most 3D models of AD show profound changes in the epidermal compartment, including hypogranulosis, spongiosis, and increased cellularity due to hyperproliferation (3D model section, Supplementary Table S3). Changes in expression of genes (detected by microarray, RNA-sequencing (RNA-Seq), or qPCR) and protein (detected by liquid chromatography mass spectrometry, Western blot, ELISA, or immunohistochemistry) can be used to evaluate disturbances in differentiation and immune response in 2D and 3D models. Lipid synthesis, which is required for optimal barrier function, can be monitored by expression of related enzymes or directly by mass spectrometry. Epidermal barrier function can be monitored in 2D and 3D models, depending on the assay. We recommend that the phenotype of any AD *in vitro* model should be extensively characterized, and should include parallel analysis of epidermal morphology, differentiation status, loss or gain of key transcripts/proteins, analysis of immune components, and assessment of functional epidermal barrier parameters. Full characterization of any AD model can inform downstream evaluation of potential therapeutic agents with respect to reversing different aspects of the disease. Testing potential targets or drugs in several model types can add rigor and indicate if a signaling pathway or protein is central to the diverse manifestations of AD.

In silico computational models

A core element of a systems biology approach is development of *in silico* computational models (mechanistic models) by integration of different types of experimental and clinical data from multiple studies, including those associated with disease conditions. *In silico* experiments, *i.e*. computer simulations or mathematical analysis of *in silico* models, can test model-specific hypotheses, predict disease prognosis or treatment outcomes, and identify knowledge gaps, guiding future experiments and clinical trials that produce further data. This iterative process refines *in silico* models, providing holistic systems-level

mechanistic insights into how perturbations (treatments or risk factors) lead to whole-organism phenotypes.

A mechanistic model describes causative interactions between the system's components involved in the phenomena of interest (*e.g.* disease or treatment outcomes). Existing mechanistic models of AD vary widely depending on the levels of interactions (tissue, cells, proteins, genes) included in the model and mathematical methods used to describe the interactions.

Domínguez-Hüttinger *et al.* developed a multi-scale deterministic model that delineates interactions between the environment, skin barrier integrity and immune activation by ordinary differential equations[54] (Table 1). Two bistable "switches" are described – the first regulating the onset of AD flares and the second controlling progression to severe and persistent disease. The model predicts, for example, that genetic predisposition to barrier dysfunction (*e.g. FLG* haploinsufficiency) predisposes to longer flares and more persistent disease and that prophylactic emollient use may be beneficial (Table 1).

Application of optimal control theory to the hybrid mathematical model can inform the design of patient-specific optimal strategies for "proactive therapy" to prevent recurrent flares once the disease has been brought under initial control [55]. For example, this computational model supports the need for higher topical steroid treatment dose after disease worsening and the potential need for more frequent than 2-3 days per week application of topical steroid treatment to maintain remission[56] in patients with *FLG* haploinsufficiency (Table 1), presenting a readily testable stratification treatment regime based on genotype.

Polak *et al.* developed a stochastic Petri net model that delineates genetic regulatory mechanisms responsible for immune responses in Langerhans cells (LCs)[57] (Table 1). The model describes reported interactions between interferon regulatory factors (IRFs), IRF transcription partners and DNA sequences in a logic-based diagram. *In vitro* experiments validated model predictions that LCs' ability to present a

peptide is altered by cytokine milieu and that a PI3Kgamma inhibitor reduces the LCs' ability to induce Th1 responses. These smaller-scale and focused mechanistic models can describe detailed interactions which are difficult to be included and validated in multi-scale models. Inclusion of the detailed interactions would make the multi-scale models too complex to interpret and to be validated, due to the current lack of quantitative dynamic data that measures the variables across different scales simultaneously.

Subramanian *et al.* used a pathway model that included manually-curated skin-specific pathways and relevant genes[58] (Table 1). Pathway enrichment analysis, using transcriptomic datasets of AD patients, provided mechanistic insights into drug actions of topical betamethasone and pimecrolimus. The pathway model would allow *in silico* experiments, once the kinetics parameters for pathways are identified, to provide quantitative and dynamic predictions of disease progression and treatment outcomes.

Population pharmacokinetic and pharmacodynamic (PK/PD) models have also been developed to describe differences and variability in pharmacological effects observed in large clinical studies for AD treatments[59] [60]. The authors identified the model parameters that can best fit to the effects of nemolizumab and dupilumab measured in terms of AD severity score or pharmacokinetics (Table 1) [59] [60]. Population PK/PD models could help achieve mechanistic understanding of pharmacological effects, if combined with mechanistic models.

One of the challenges in developing mechanistic models is identification of the components and the pathways that are relevant to the model-specific hypothesis to be tested. This can be achieved by unbiased multivariate analyses of a collection of large-scale data, for example by machine learning data analysis. Application of machine learning methods to AD-related data is relatively limited at present, but some relevant works have been already published. Thijs *et al.* developed a piecewise linear mixed model to predict AD severity scores after different treatments[61] and Kiiski *et al*. developed a multivariate logistic regression model to predict a "good treatment response" [62]. A sufficient level of cross-validation is crucial

to reduce bias and to ensure the general applicability of models that have predictive power beyond mere description of data.

All the models presented above were developed based on the published data derived from studies in which the inclusion and exclusion criteria for AD were specified. Whilst the majority of studies utilised the Hanifin and Rajka criteria and specified further clinical (including co-morbidities) and demographical details, it is clear that patients with AD present with a wide spectrum of clinical and molecular features (including for example a greater heterogeneity in transcriptomic profile of lesional skin compared to psoriasis)[63].

Future developments

The development of more sophisticated human models of AD that integrate large scale clinical and 'omic' data offer the potential for a deeper understanding of disease endotypes, molecular mechanisms underlying key pathogenic events and clinical hallmarks of AD, as well as prediction of therapeutic outcomes, including comorbidity at the level of an individual patient. Accepting that, by definition, these human models are based upon a reductionist approach, they need to reflect the complexity of AD pathogenesis, including epidermal barrier dysfunction, altered penetration of chemicals and allergens, host/environment interaction, type 2 immunity, and tissue remodeling. We have illustrated in this review that the main approaches available today are *in vitro* models, identification and characterization of human inherited syndromes resembling AD, *in vivo* challenges of AD patients, as well as *in silico* models. Here, we speculate how the future of AD research will likely inform the development of more refined human models of AD.

Refinement is likely to depend, at least in part, upon methodological advances in the field and the additional information generated by novel approaches. For example, single cell sequencing has recently identified novel rare but important immunological subsets[64] and intravital photon microscopy has enabled visualization of cell-cell communication during inflammation[65 66]. Application of this technology to AD is likely to inform the inclusion of distinct epithelial and immune cell types[64] and/or genetically modified

primary human cells[67]. Furthermore, small-scale spheroid organoids may enhance high-throughput approaches in the field[68]. Finally, we expect that a technological breakthrough in the development of three-dimensional skin models will be facilitated by cell printers[69, 70].

Deep neural networks are being applied as artificial intelligence tools to facilitate physician interpretation in the field of melanoma diagnostics [71] and increasingly as methods to enable large data set integration. The first examples of disease classifiers[72] and prediction of disease severity from biomarker sets[61, 73, 74] have recently been published, and we expect this line of development to continue while ensuring a sufficient level of validation. We anticipate that refinement of these methods, in combination with *in silico* models, may lead to computational approaches and predictive models applied to diagnostics and therapeutic stratification. The descriptive disease ontology of inflammatory skin diseases will need to be revised by shifting to pathogenesis-oriented structure[75] and, in the future, by better definition of disease endotypes based on integration of multiomics data, clinical features, and clinical response to therapy in light of *in silico* models as assessed in large-scale and longitudinal cohorts[76]. These advances are likely to inform the development of many of the current models.

To achieve a substantial breakthrough, though, we expect that different approaches will need to be combined, integrated, standardized, and performed at larger scale (Figure 3). For example, observations made in rare human disease variants or by specific challenge models in AD patients may be validated *in vitro* and mapped to disease signatures *in silico*. Validation of functional hypotheses will increasingly depend upon cross-referencing of data derived from clinical samples with outputs from *in vitro* models. Integration of clinical, biomarker, PK/PD (topical and/or systemic) and clinical outcome data will inform therapy development and precision medicine. Notably, all of our models depend on how precisely a particular question is asked and the quality of the clinical input, including the clinical metadata and integration with omics data derived from clinical samples. Finally, advanced statistical and machine learning analysis combined with *in silico* predictive modelling will be required to integrate information

345 throughout all described layers and data sets to elucidate underlying mechanisms (and endotypes), further

346 highlighting the importance of data standardization and scientific networking.

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Figure legends

Figure 1. Diagrammatic representation of 'Human knockout' monogenic models providing insight into the pathomechanisms of AD. Specific genetic variants affecting the structural and/or immune functions of skin or other organs recapitulate features, but not the entire phenotype, of atopic inflammation and AD. *CARD11*, caspase recruitment domain-containing protein 11; *CDSN*, corneodesmosin; *CTLA4*, cytotoxic T lymphocyte-associated protein 4; *DOCK8*, dedicator of cytokinesis 8; *DSG1*, desmoglein 1; *DSP*, desmoplakin; *FLG*, filaggrin; *FOXP3*, forkhead-box-protein 3; *IL2RA,* interleukin-2 receptor alpha; *IL4RA*, interleukin 4 receptor alpha; *IFNGR1*, interferon gamma receptor 1; *MALT1*, mucosa-associated lymphoid tissue lymphoma translocation protein 1; *PGM3*, phosphoglucomutase 3; *RAG1, RAG2,* recombination-activated gene 1 and 2; *SPINK5*, serine protease inhibitor Kazal type 5; *STAT3*, signal transducer and activator of transcription 3.

Figure 2. Human *in vitro* models of AD. *In vitro* models can be designed to address specific experimental questions based on the input materials of the cultures. Assessment of the cultures, or output, depends on the type of culture. HEE, human epidermal equivalent; HSE, human skin equivalent (inset: fibroblasts in collagen); *FLG,* filaggrin; *IVL*, involucrin; *KRT10*, keratin 10; *DSG1*, desmoglein 1; *CDSN*, corneodesmosin; *TSLP*, thymic stromal lymphopoietin; TEER, trans-epithelial electrical resistance.

Figure 3. Interconnected multi-layer networks: the future of human AD modelling. To answer clinically relevant questions such as identification of distinct disease endotypes, elucidation of molecular pathomechanisms, or prediction of therapeutic response, a combination of innovative *in vitro* and *in silico* models obtained by a systems biology approach and machine learning algorithms will be needed.

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Table 1

Model Type	Scientific Merits	Clinical Utility	Limitations	Key Features	Key Findings/Predictions	Refs
Multi-scale mechanistic model	Mechanistic understanding of system-level effects of potential triggers and processes on disease state	Identification of therapeutic targets, and their mechanisms, for further clinical investigation. Prediction of dynamic effects of therapeutics, leading to patient stratification	Models developed based on hypothesized relationships that were previously described experimentally.	A hybrid ordinary differential equation model of the dynamic interplay between skin barrier function, immune responses and environmental stressors that determines AD pathogenesis	Preventive effects of emollients against AD progression (shown by clinical trials). Synergistic effects of environmental (eg. microbiome) and genetic (eg. FLG) risk factors on AD progression (shown by mice experiments with ovalbumin challenge or dose-dependent effects of FLG deficiency)	[54]
				A hybrid model of treatment effects of corticosteroids and emollients on AD pathogenesis and exploration of optimal regimes for induction of remission and maintance of remission	Poor adherence to the suggested optimal treatment schedule leads to higher treatment doses. Application of corticosteroids for 2 consecutive days per week is optimal for maintenance period	[55]
Gene regulatory network model	Understanding of gene regulatory mechanisms behind disease processes	Identification of therapeutic targets, and their mechanisms, at the gene regulation level.	Models developed based on published genetic interactions.	Stochastic Petri Net model of Interferon regulatory factors gene regulatory network in response to in *vitro* treatment of Langerhans cells (LC) with TNFα and TSLP	In *vitro* experiments validated predictions that LCs' ability to present a peptide is altered by cytokine milieu and that PI3Kg inhibitor reduces the LC's ability to induce Th1 responses	[57]
Pathway models	Understanding of disease mechanisms	Identification of therapeutic targets, and their mechanisms	Models developed based on published pathways.	A pathway model including 35 manually-curated skin-specific pathways and 2600+ genes.	Pathway enrichment analysis using transcriptomic datasets of 10 AD patients treated with betamethasone valerate and pimecrolimus predicted mechanism of action of both drugs on human skin	[58]
Population PK/PD models	Understanding of differences and variability in pharmacological effects among a target population from clinical trials data	Prediction of optimal dose regimen. Testing effects of weight, gender etc.	Requires a large clinical data to have sufficient predictive power	PK/PD model for serum nemolizumab and pruritus VAS developed from 299 patients' time course data	An appropriate flat dose regimen that is independent of body weights	[59]
				Two compartment PK model for dupilumab developed from data of 197 healthy volunteers and AD patients from 6 studies	Production rate of IL4Ra is similar for AD patients and normal volunteers, and does not change over time	[60]
Machine learning predictive models	Unbiased analyses of differences between disease and non-disease (including treated) tissue/ patients and prediction of clinical outcomes (prognostic and therapeutic)	Identification of disease and therapeutic targets. Findings can feed into mechanistic models	Causative mechanisms remain largely unknown. Machine learning applications to atopic eczema relatively limited at present	Piecewise linear mixed models to predict EASI scores at 3 future timepoints from baseline biomarkers. Developed from data of 150 serum biomarkers measured in 193 AD patients	Combination of TARC, IL-22 and sIL-2R provides a good predictor for future EASI	[61]
				Multivariate logistic regression model to identify predictors of long-term response to topical maintenance treatment in AD on 169 patients.	Serum total IgE (rather than the initial severity) is the most important factor predicting a good long-term treatment outcome	[62]

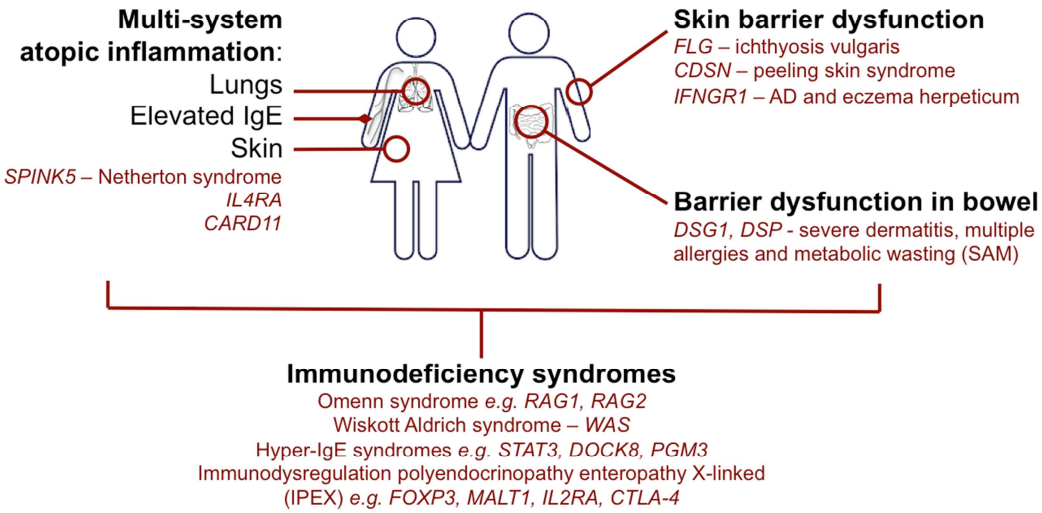
References

1. Paternoster L, Standl M, Waage J, Baurecht H, Hotze M, Strachan DP, et al. Multi-ancestry genome-wide association study of 21,000 cases and 95,000 controls identifies new risk loci for atopic dermatitis. Nat Genet 2015; 47:1449-56.
2. Weidinger S, Novak N. Atopic dermatitis. Lancet 2016; 387:1109-22.
3. Brunner PM, Silverberg JI, Guttman-Yassky E, Paller AS, Kabashima K, Amagai M, et al. Increasing Comorbidities Suggest that Atopic Dermatitis Is a Systemic Disorder. J Invest Dermatol 2017; 137:18-25.
4. Jin H, He R, Oyoshi M, Geha RS. Animal models of atopic dermatitis. J Invest Dermatol 2009; 129:31-40.
5. Ewald DA, Noda S, Oliva M, Litman T, Nakajima S, Li X, et al. Major differences between human atopic dermatitis and murine models, as determined by using global transcriptomic profiling. J Allergy Clin Immunol 2017; 139:562-71.
6. Irvine AD, McLean WH, Leung DY. Filaggrin mutations associated with skin and allergic diseases. N Engl J Med 2011; 365:1315-27.
7. Barzaghi F, Amaya Hernandez LC, Neven B, Ricci S, Kucuk ZY, Bleesing JJ, et al. Long-term follow-up of IPEX syndrome patients after different therapeutic strategies: An international multicenter retrospective study. J Allergy Clin Immunol 2018; 141:1036-49 e5.
8. Abolhassani H, Chou J, Bainter W, Platt CD, Tavassoli M, Momen T, et al. Clinical, immunologic, and genetic spectrum of 696 patients with combined immunodeficiency. J Allergy Clin Immunol 2018; 141:1450-8.
9. Boos AC, Hagl B, Schlesinger A, Halm BE, Ballenberger N, Pinarci M, et al. Atopic dermatitis, STAT3- and DOCK8-hyper-IgE syndromes differ in IgE-based sensitization pattern. Allergy 2014; 69:943-53.
10. Turjanmaa K, Darsow U, Niggemann B, Rance F, Vanto T, Werfel T. EAACI/GA2LEN position paper: present status of the atopy patch test. Allergy 2006; 61:1377-84.
11. Werfel T, Allam JP, Biedermann T, Eyerich K, Gilles S, Guttman-Yassky E, et al. Cellular and molecular immunologic mechanisms in patients with atopic dermatitis. J Allergy Clin Immunol 2016; 138:336-49.
12. Darsow U, Vieluf D, Ring J. Evaluating the relevance of aeroallergen sensitization in atopic eczema with the atopy patch test: a randomized, double-blind multicenter study. Atopy Patch Test Study Group. J Am Acad Dermatol 1999; 40:187-93.
13. Schmid-Grendelmeier P, Fluckiger S, Disch R, Trautmann A, Wuthrich B, Blaser K, et al. IgE-mediated and T cell-mediated autoimmunity against manganese superoxide dismutase in atopic dermatitis. J Allergy Clin Immunol 2005; 115:1068-75.
14. Ungar B, Correa da Rosa J, Shemer A, Czarnowicki T, Estrada YD, Fuentes-Duculan J, et al. Patch testing of food allergens promotes Th17 and Th2 responses with increased IL-33: a pilot study. Exp Dermatol 2017; 26:272-5.
15. Darsow U, Vieluf D, Ring J. Atopy patch test with different vehicles and allergen concentrations: an approach to standardization. J Allergy Clin Immunol 1995; 95:677-84.
16. Sager N, Feldmann A, Schilling G, Kreitsch P, Neumann C. House dust mite-specific T cells in the skin of subjects with atopic dermatitis: frequency and lymphokine profile in the allergen patch test. J Allergy Clin Immunol 1992; 89:801-10.
17. Gittler JK, Shemer A, Suarez-Farinas M, Fuentes-Duculan J, Gulewicz KJ, Wang CQ, et al. Progressive activation of T(H)2/T(H)22 cytokines and selective epidermal proteins characterizes acute and chronic atopic dermatitis. J Allergy Clin Immunol 2012; 130:1344-54.
18. Kerschenlohr K, Decard S, Przybilla B, Wollenberg A. Atopy patch test reactions show a rapid influx of inflammatory dendritic epidermal cells in patients with extrinsic atopic dermatitis and patients with intrinsic atopic dermatitis. J Allergy Clin Immunol 2003; 111:869-74.
19. Guttman-Yassky E, Nograles KE, Krueger JG. Contrasting pathogenesis of atopic dermatitis and psoriasis--part II: immune cell subsets and therapeutic concepts. J Allergy Clin Immunol 2011; 127:1420-32.

20. Eyerich S, Onken AT, Weidinger S, Franke A, Nasorri F, Pennino D, et al. Mutual antagonism of T cells causing psoriasis and atopic eczema. N Engl J Med 2011; 365:231-8.
21. Eyerich K, Pennino D, Scarponi C, Foerster S, Nasorri F, Behrendt H, et al. IL-17 in atopic eczema: linking allergen-specific adaptive and microbial-triggered innate immune response. J Allergy Clin Immunol 2009; 123:59-66 e4.
22. Niebuhr M, Scharonow H, Gathmann M, Mamerow D, Werfel T. Staphylococcal exotoxins are strong inducers of IL-22: A potential role in atopic dermatitis. J Allergy Clin Immunol 2010; 126:1176-83 e4.
23. Langer K, Breuer K, Kapp A, Werfel T. Staphylococcus aureus-derived enterotoxins enhance house dust mite-induced patch test reactions in atopic dermatitis. Exp Dermatol 2007; 16:124-9.
24. Hamann CR, Hamann D, Egeberg A, Johansen JD, Silverberg J, Thyssen JP. Association between atopic dermatitis and contact sensitization: A systematic review and meta-analysis. Journal of the American Academy of Dermatology 2017; 77:70-8.
25. Correa da Rosa J, Malajian D, Shemer A, Rozenblit M, Dhingra N, Czarnowicki T, et al. Patients with atopic dermatitis have attenuated and distinct contact hypersensitivity responses to common allergens in skin. J Allergy Clin Immunol 2015; 135:712-20.
26. Dhingra N, Shemer A, Correa da Rosa J, Rozenblit M, Fuentes-Duculan J, Gittler JK, et al. Molecular profiling of contact dermatitis skin identifies allergen-dependent differences in immune response. J Allergy Clin Immunol 2014; 134:362-72.
27. Garzorz-Stark N, Lauffer F, Krause L, Thomas J, Atenhan A, Franz R, et al. Toll-like receptor 7/8 agonists stimulate plasmacytoid dendritic cells to initiate TH17-deviated acute contact dermatitis in human subjects. J Allergy Clin Immunol 2018; 141:1320-33 e11.
28. Newell L, Polak ME, Perera J, Owen C, Boyd P, Pickard C, et al. Sensitization via healthy skin programs Th2 responses in individuals with atopic dermatitis. J Invest Dermatol 2013; 133:2372-80.
29. Werfel T, Heratizadeh A, Niebuhr M, Kapp A, Roesner LM, Karch A, et al. Exacerbation of atopic dermatitis on grass pollen exposure in an environmental challenge chamber. J Allergy Clin Immunol 2015; 136:96-103 e9.
30. Engebretsen KA, Bager P, Wohlfahrt J, Skov L, Zachariae C, Nybo Andersen AM, et al. Prevalence of atopic dermatitis in infants by domestic water hardness and season of birth: Cohort study. J Allergy Clin Immunol 2017; 139:1568-74 e1.
31. Engebretsen KA, Kezic S, Jakasa I, Hedengran A, Linneberg A, Skov L, et al. Effect of atopic skin stressors on natural moisturizing factors and cytokines in healthy adult epidermis. Br J Dermatol 2018.
32. Guttman-Yassky E, Ungar B, Malik K, Dickstein D, Suprun M, Estrada YD, et al. Molecular signatures order the potency of topically applied anti-inflammatory drugs in patients with atopic dermatitis. J Allergy Clin Immunol 2017; 140:1032-42 e13.
33. Otsuka A, Doi H, Egawa G, Maekawa A, Fujita T, Nakamizo S, et al. Possible new therapeutic strategy to regulate atopic dermatitis through upregulating filaggrin expression. J Allergy Clin Immunol 2014; 133:139-46.e1-10.
34. Howell MD, Kim BE, Gao P, Grant AV, Boguniewicz M, DeBenedetto A, et al. Cytokine modulation of atopic dermatitis filaggrin skin expression. J Allergy Clin Immunol 2009; 124:R7-r12.
35. Kamsteeg M, Bergers M, de Boer R, Zeeuwen PL, Hato SV, Schalkwijk J, et al. Type 2 helper T-cell cytokines induce morphologic and molecular characteristics of atopic dermatitis in human skin equivalent. Am J Pathol 2011; 178:2091-9.
36. Yuki T, Tobiishi M, Kusaka-Kikushima A, Ota Y, Tokura Y. Impaired Tight Junctions in Atopic Dermatitis Skin and in a Skin-Equivalent Model Treated with Interleukin-17. PLoS One 2016; 11:e0161759.
37. Hanel KH, Pfaff CM, Cornelissen C, Amann PM, Marquardt Y, Czaja K, et al. Control of the Physical and Antimicrobial Skin Barrier by an IL-31-IL-1 Signaling Network. J Immunol 2016; 196:3233-44.
38. Rouaud-Tinguely P, Boudier D, Marchand L, Barruche V, Bordes S, Coppin H, et al. From the morphological to the transcriptomic characterization of a compromised three-dimensional in vitro model mimicking atopic dermatitis. Br J Dermatol 2015; 173:1006-14.

39. De Vuyst E, Giltaire S, Lambert de Rouvroit C, Malaisse J, Mound A, Bourtembourg M, et al. Methyl-beta-cyclodextrin concurs with interleukin (IL)-4, IL-13 and IL-25 to induce alterations reminiscent of atopic dermatitis in reconstructed human epidermis. Exp Dermatol 2018; 27:435-7.
40. Danso MO, van Drongelen V, Mulder A, van Esch J, Scott H, van Smeden J, et al. TNF-alpha and Th2 cytokines induce atopic dermatitis-like features on epidermal differentiation proteins and stratum corneum lipids in human skin equivalents. J Invest Dermatol 2014; 134:1941-50.
41. Nygaard U, van den Bogaard EH, Niehues H, Hvid M, Deleuran M, Johansen C, et al. The "Alarmins" HMBG1 and IL-33 Downregulate Structural Skin Barrier Proteins and Impair Epidermal Growth. Acta Derm Venereol 2017; 97:305-12.
42. Elias MS, Long HA, Newman CF, Wilson PA, West A, McGill PJ, et al. Proteomic analysis of filaggrin deficiency identifies molecular signatures characteristic of atopic eczema. J Allergy Clin Immunol 2017; 140:1299-309.
43. Honzke S, Wallmeyer L, Ostrowski A, Radbruch M, Mundhenk L, Schafer-Korting M, et al. Influence of Th2 Cytokines on the Cornified Envelope, Tight Junction Proteins, and ss-Defensins in Filaggrin-Deficient Skin Equivalents. J Invest Dermatol 2016; 136:631-9.
44. Pastore S, Fanales-Belasio E, Albanesi C, Chinni LM, Giannetti A, Girolomoni G. Granulocyte macrophage colony-stimulating factor is overproduced by keratinocytes in atopic dermatitis. Implications for sustained dendritic cell activation in the skin. J Clin Invest 1997; 99:3009-17.
45. van Drongelen V, Danso MO, Out JJ, Mulder A, Lavrijsen AP, Bouwstra JA, et al. Explant cultures of atopic dermatitis biopsies maintain their epidermal characteristics in vitro. Cell Tissue Res 2015; 361:789-97.
46. Bogiatzi SI, Fernandez I, Bichet JC, Marloie-Provost MA, Volpe E, Sastre X, et al. Cutting Edge: Proinflammatory and Th2 cytokines synergize to induce thymic stromal lymphopoietin production by human skin keratinocytes. J Immunol 2007; 178:3373-7.
47. Berroth A, Kuhnl J, Kurschat N, Schwarz A, Stab F, Schwarz T, et al. Role of fibroblasts in the pathogenesis of atopic dermatitis. J Allergy Clin Immunol 2013; 131:1547-54.
48. Wallmeyer L, Dietert K, Sochorova M, Gruber AD, Kleuser B, Vavrova K, et al. TSLP is a direct trigger for T cell migration in filaggrin-deficient skin equivalents. Sci Rep 2017; 7:774.
49. Lecas S, Boursier E, Fitoussi R, Vie K, Momas I, Seta N, et al. In vitro model adapted to the study of skin ageing induced by air pollution. Toxicol Lett 2016; 259:60-8.
50. Maboni G, Davenport R, Sessford K, Baiker K, Jensen TK, Blanchard AM, et al. A Novel 3D Skin Explant Model to Study Anaerobic Bacterial Infection. Front Cell Infect Microbiol 2017; 7:404.
51. Marionnet C, Pierrard C, Lejeune F, Sok J, Thomas M, Bernerd F. Different oxidative stress response in keratinocytes and fibroblasts of reconstructed skin exposed to non extreme daily-ultraviolet radiation. PLoS One 2010; 5:e12059.
52. Abaci HE, Guo Z, Doucet Y, Jackow J, Christiano A. Next generation human skin constructs as advanced tools for drug development. Exp Biol Med (Maywood) 2017; 242:1657-68.
53. Castex-Rizzi N, Galliano MF, Aries MF, Hernandez-Pigeon H, Vaissiere C, Delga H, et al. In vitro approaches to pharmacological screening in the field of atopic dermatitis. Br J Dermatol 2014; 170 Suppl 1:12-8.
54. Dominguez-Huttinger E, Christodoulides P, Miyauchi K, Irvine AD, Okada-Hatakeyama M, Kubo M, et al. Mathematical modeling of atopic dermatitis reveals "double-switch" mechanisms underlying 4 common disease phenotypes. J Allergy Clin Immunol 2017; 139:1861-72.e7.
55. Christodoulides P, Hirata Y, Dominguez-Huttinger E, Danby SG, Cork MJ, Williams HC, et al. Computational design of treatment strategies for proactive therapy on atopic dermatitis using optimal control theory. Philos Trans A Math Phys Eng Sci 2017; 375.
56. Schmitt J, von Kobyletzki L, Svensson A, Apfelbacher C. Efficacy and tolerability of proactive treatment with topical corticosteroids and calcineurin inhibitors for atopic eczema: systematic review and meta-analysis of randomized controlled trials. Br J Dermatol 2011; 164:415-28.
57. Polak ME, Ung CY, Masapust J, Freeman TC, Ardern-Jones MR. Petri Net computational modelling of Langerhans cell Interferon Regulatory Factor Network predicts their role in T cell activation. Sci Rep 2017; 7:668.

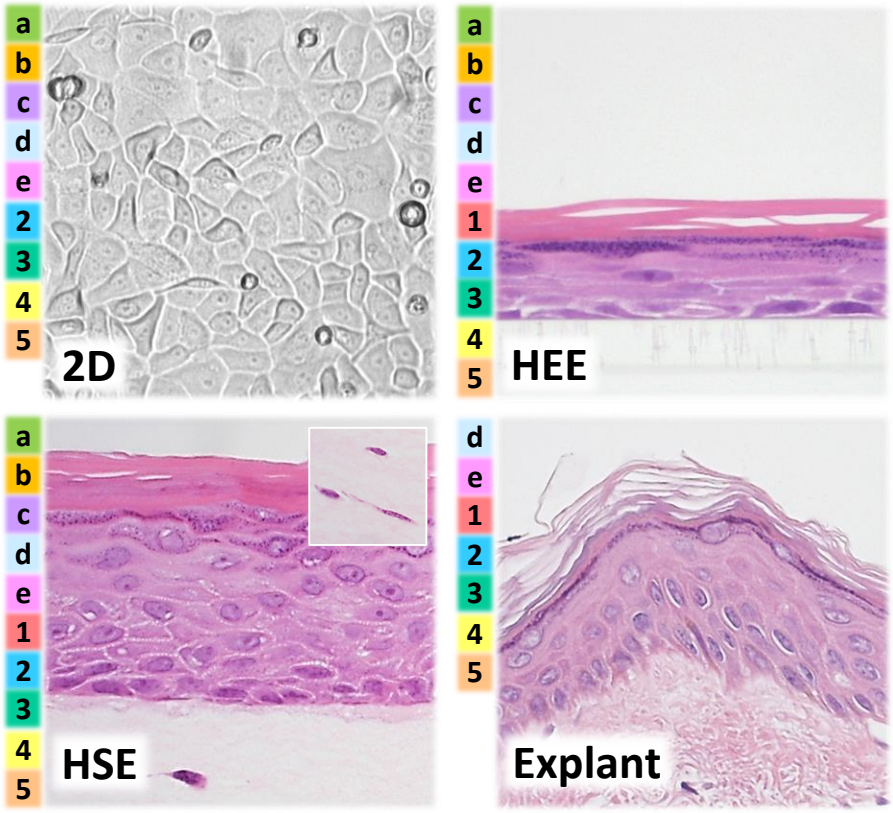
58. Subramanian I, Singh VK, Jere A. Elucidating mechanistic insights into drug action for atopic dermatitis: a systems biology approach. BMC Dermatol 2018; 18:3.
59. Saito T, Iida S, Terao K, Kumagai Y. Dosage Optimization of Nemolizumab Using Population Pharmacokinetic and Pharmacokinetic-Pharmacodynamic Modeling and Simulation. J Clin Pharmacol 2017; 57:1564-72.
60. Kovalenko P, DiCioccio AT, Davis JD, Li M, Ardeleanu M, Graham N, et al. Exploratory Population PK Analysis of Dupilumab, a Fully Human Monoclonal Antibody Against IL-4Ralpha, in Atopic Dermatitis Patients and Normal Volunteers. CPT Pharmacometrics Syst Pharmacol 2016; 5:617-24.
61. Thijs JL, Drylewicz J, Fiechter R, Strickland I, Sleeman MA, Herath A, et al. EASI p-EASI: Utilizing a combination of serum biomarkers offers an objective measurement tool for disease severity in atopic dermatitis patients. J Allergy Clin Immunol 2017; 140:1703-5.
62. Kiiski V, Karlsson O, Remitz A, Reitamo S. High Serum Total IgE Predicts Poor Long-term Outcome in Atopic Dermatitis. Acta Dermato Venereologica 2015; 95:943-7.
63. Guttman-Yassky E, Krueger JG. Atopic dermatitis and psoriasis: two different immune diseases or one spectrum? Curr Opin Immunol 2017; 48:68-73.
64. Villani AC, Satija R, Reynolds G, Sarkizova S, Shekhar K, Fletcher J, et al. Single-cell RNA-seq reveals new types of human blood dendritic cells, monocytes, and progenitors. Science 2017; 356.
65. Reber LL, Sibilano R, Starkl P, Roers A, Grimbaldeston MA, Tsai M, et al. Imaging protective mast cells in living mice during severe contact hypersensitivity. JCI Insight 2017; 2.
66. Dudeck J, Medyukhina A, Frobel J, Svensson CM, Kotrba J, Gerlach M, et al. Mast cells acquire MHCII from dendritic cells during skin inflammation. J Exp Med 2017; 214:3791-811.
67. Niehues H, Bouwstra JA, El Ghalbzouri A, Brandner JM, Zeeuwen P, van den Bogaard EH. 3D skin models for 3R research: The potential of 3D reconstructed skin models to study skin barrier function. Exp Dermatol 2018; 27:501-11.
68. Lauffer F, Jargosch M, Krause L, Garzorz-Stark N, Franz R, Roenneberg S, et al. Type I Immune Response Induces Keratinocyte Necroptosis and Is Associated with Interface Dermatitis. J Invest Dermatol 2018.
69. Kim BS, Kwon YW, Kong JS, Park GT, Gao G, Han W, et al. 3D cell printing of in vitro stabilized skin model and in vivo pre-vascularized skin patch using tissue-specific extracellular matrix bioink: A step towards advanced skin tissue engineering. Biomaterials 2018; 168:38-53.
70. Pourchet LJ, Thepot A, Albouy M, Courtial EJ, Boher A, Blum LJ, et al. Human Skin 3D Bioprinting Using Scaffold-Free Approach. Adv Healthc Mater 2017; 6.
71. Esteva A, Kuprel B, Novoa RA, Ko J, Swetter SM, Blau HM, et al. Dermatologist-level classification of skin cancer with deep neural networks. Nature 2017; 542:115-8.
72. Quaranta M, Knapp B, Garzorz N, Mattii M, Pullabhatla V, Pennino D, et al. Intraindividual genome expression analysis reveals a specific molecular signature of psoriasis and eczema. Sci Transl Med 2014; 6:244ra90.
73. Ungar B, Garcet S, Gonzalez J, Dhingra N, Correa da Rosa J, Shemer A, et al. An Integrated Model of Atopic Dermatitis Biomarkers Highlights the Systemic Nature of the Disease. J Invest Dermatol 2017; 137:603-13.
74. Krause L, Mourantchanian V, Brockow K, Theis FJ, Schmidt-Weber CB, Knapp B, et al. A computational model to predict severity of atopic eczema from 30 serum proteins. J Allergy Clin Immunol 2016; 138:1207-10 e2.
75. Eyerich K, Eyerich S. Immune response patterns in non-communicable inflammatory skin diseases. J Eur Acad Dermatol Venereol 2018; 32:692-703.
76. Paternoster L, Savenije OEM, Heron J, Evans DM, Vonk JM, Brunekreef B, et al. Identification of atopic dermatitis subgroups in children from 2 longitudinal birth cohorts. J Allergy Clin Immunol 2018; 141:964-71.



Input

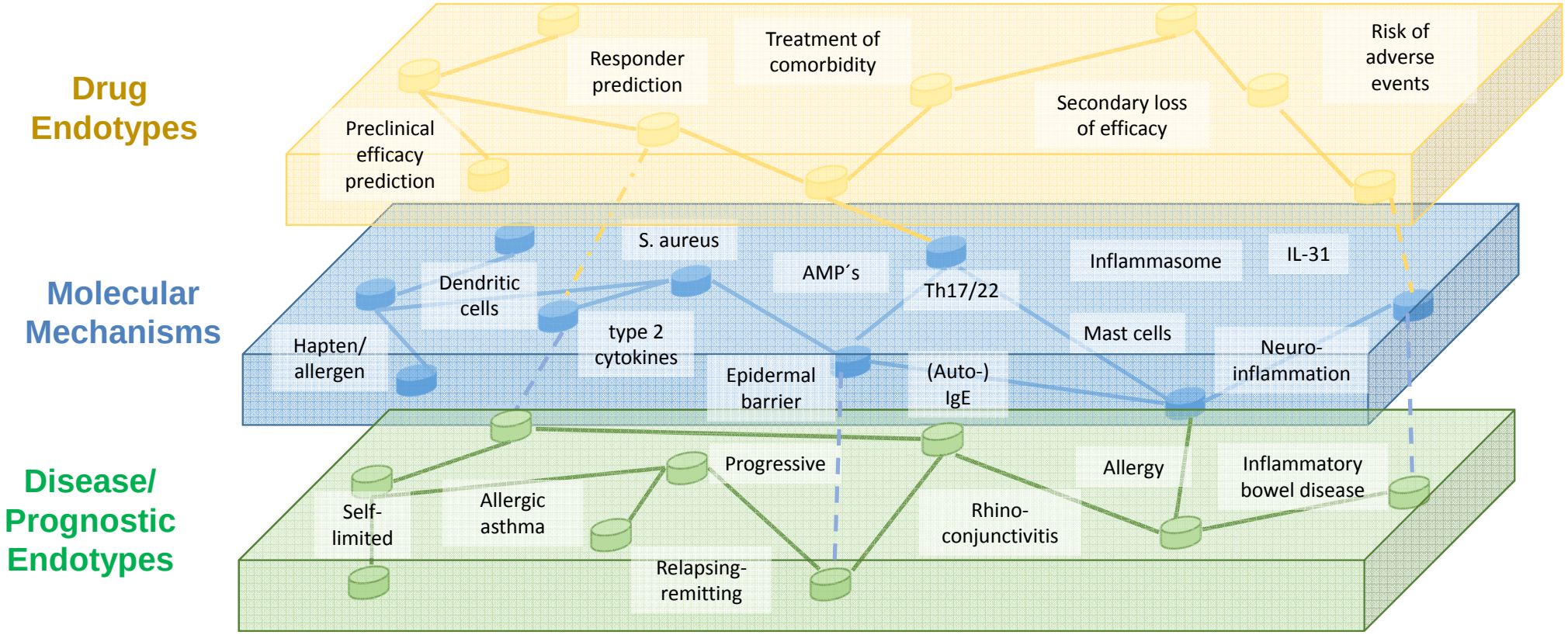
- **a** Normal Keratinocytes: *primary/immortalized*
- **b** Gene (e.g., *FLG*) knockdown: *siRNA/shRNA/CRISPR*
- **c** Co-Culture: *T-cells, neurons, fibroblasts*
- **d** Patient-derived tissue/cells: *immune, keratinocytes, etc.*
- **e** Cytokine treatment: *interleukin(s)*

In Vitro Model



Output

- **1** Epidermal Morphology: *e.g., spongiosis*
- **2** Differentiation: *FLG, IVL, KRT10, LOR, DSG1, CDSN, etc.*
- **3** Keratinocyte / immune cell factors: *TSLP, Th2 cytokines*
- **4** Barrier Function: *Lipid profile, TEER, permeability*
- **5** Key Transcript and Protein Changes *immune, barrier, etc.*



Integration of data by systems biology and machine learning

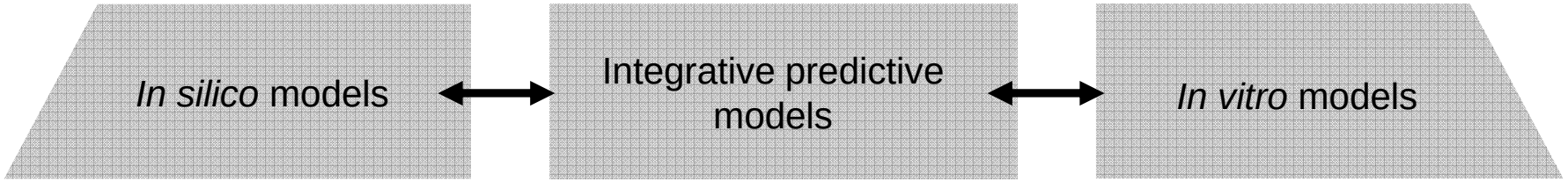


Table 1. Genetic disease models of AD

Genetic disease	Gene and mutation type(s)	Phenotype(s)	Mechanistic insights	Clinical utility	Limitations	Pathway relevance for drug development	Refs.
Skin barrier dysfunction							
Ichthyosis vulgaris (IV)	*FLG* Loss of function mutations semi-dominant in IV and complex trait in AD	Early onset, severe and persistent AD with & without other atopic diseases; predisposition to eczema herpeticum (EH)	Understanding that skin barrier dysfunction predates atopic inflammation	Illustrates importance of barrier repair	Molecular mechanisms and control pathways remain unclear		1, 2
Generalised peeling skin	*CDSN* Loss of function mutation autosomal recessive	Ichthyosiform erythroderma, pruritus and food allergies	Confirms the role of corneodesmosin in epidermal adhesion	Understanding that skin barrier dysfunction predates atopic inflammation			3
AD and eczema herpeticum	*IFNGR1* Loss of function mutation Complex trait	AD and eczema herpeticum (EH)	Defective systemic IFN-gamma immune response accounts for disseminated viral skin infections	Helps to explain why a subset of AD patients suffer recurrent EH	Does not explain all cases of EH		4
Netherton syndrome	*SPINK5* Loss of function mutation Autosomal recessive	Congenital ichthyosis, severe atopic disease, elevated IgE, hypereosinophilia, infections	Single nucleotide variants associated with AD. Illustrates role of epidermal protease inhibitors and kallikrein proteases in regulating epidermal barrier function	Understanding that skin barrier dysfunction predates atopic inflammation		Protease inhibitors	5
Systemic atopic inflammation							
Atopic disease	*IL4RA* Gain of function Complex trait	Elevated IgE with & without AD	Mutation found in severe cases is also a common risk allele in the population	Evidence of role for IL-4 in atopic inflammation		IL-4RA	6
Severe atopic disease	*CARD11* Heterozygous mutations Loss of function and dominant negative effect	Severe AD with & without infection	Illustrates importance of lymphocyte receptor signalling	mTORC1 and IFN-gamma production defects can be partially rescued by glutamine supplementation	Unclear whether this mechanism plays a role in prevalent AD	NFKB and MALT1	7
Skin inflammation and gastrointestinal inflammation							
SAM (Severe dermatitis, multiple Allergies and Metabolic wasting)	*DSG1* Homozygous loss of function mutations	Ichthyosiform erythroderma, atopic disease and failure to thrive	*DSG1* mutations lead to loss of cell-cell adhesion in epidermis	Structural epidermal defects lead to atopic inflammation			8
SAM	*DSP* Heterozygous mutation	Ichthyosiform erythroderma, atopic disease and failure to thrive	*DSP* mutations result in disrupted keratin filament attachment to desmosomes	Structural epidermal defects lead to atopic inflammation	Other *DSP* mutations cause different phenotypes without atopic manifestations		9
Immunodeficiency syndromes							
Hyper-IgE	*STAT3* Dominant negative mutations	AD-like skin inflammation, elevated IgE, immunodeficiency leading to infection	Illustrates role of STAT3 in signal transduction for multiple cytokines	Biologic treatments targeting IgE have limited clinical efficacy for AD	Immunodeficiency is not a prominent feature of AD	STAT6: downstream of JAKs in Th2 inflammation	10
	DOCK8 Autosomal recessive loss of function mutations	AD-like skin inflammation, elevated IgE, immunodeficiency leading to infection	Aberrations of T cell and NK cell migration to skin can cause atopic inflammation	Antiviral and antibacterial prophylaxis, immunoglobulin replacement and HSCT			11
Omenn syndrome	Hypomorphic missense mutations in a range of genes involved T and B cell development *eg. RAG1, RAG2*	AD-like skin inflammation, elevated IgE, immunodeficiency leading to infection	Skin inflammation can occur in the absence of adaptive immunity, also seen in mice				12
Hyper-IgE like syndrome	*PGM3* Autosomal recessive loss of function mutations	AD-like skin inflammation, atopy, immune deficiency, autoimmunity and neurocognitive impairment	Role of glycosylation in immune regulation and systemic atopy				13
Wiskott-Aldrich	*WAS* X-linked mutations	AD-like skin inflammation, severe immunodeficiency, autoimmunity and malignancy	Systemic imbalances in Treg populations can drive cutaneous AD like inflammation	Requires HSCT		OX40	14
IPEX and IPEX-like syndromes	*FOXP3, MALT1, IL2RA, CTLA-4* Autosomal recessive	Immune dysregulation, polyendocrinopathy, enteropathy and AD-like skin inflammation	Role of autoimmunity in AD-like inflammation	Immunosuppressive treatment or HSCT		*FOXP3* as possible target for gene editing	15, 16

	mutations						

References

1. Gao PS, Rafaels NM, Hand T, Murray T, Boguniewicz M, Hata T, et al. Filaggrin mutations that confer risk of atopic dermatitis confer greater risk for eczema herpeticum. J Allergy Clin Immunol 2009; 124:507-13, 13 e1-7.
2. Palmer CN, Irvine AD, Terron-Kwiatkowski A, Zhao Y, Liao H, Lee SP, et al. Common loss-of-function variants of the epidermal barrier protein filaggrin are a major predisposing factor for atopic dermatitis. Nat Genet 2006; 38:441-6.
3. Oji V, Eckl KM, Aufenvenne K, Natebus M, Tarinski T, Ackermann K, et al. Loss of corneodesmosin leads to severe skin barrier defect, pruritus, and atopy: unraveling the peeling skin disease. Am J Hum Genet 2010; 87:274-81.
4. Gao L, Bin L, Rafaels NM, Huang L, Potee J, Ruczinski I, et al. Targeted deep sequencing identifies rare loss-of-function variants in IFNGR1 for risk of atopic dermatitis complicated by eczema herpeticum. J Allergy Clin Immunol 2015; 136:1591-600.
5. Chavanas S, Bodemer C, Rochat A, Hamel-Teillac D, Ali M, Irvine AD, et al. Mutations in SPINK5, encoding a serine protease inhibitor, cause Netherton syndrome. Nat Genet 2000; 25:141-2.
6. Hershey GK, Friedrich MF, Esswein LA, Thomas ML, Chatila TA. The association of atopy with a gain-of-function mutation in the alpha subunit of the interleukin-4 receptor. N Engl J Med 1997; 337:1720-5.
7. Ma CA, Stinson JR, Zhang Y, Abbott JK, Weinreich MA, Hauk PJ, et al. Germline hypomorphic CARD11 mutations in severe atopic disease. Nat Genet 2017; 49:1192-201.
8. Samuelov L, Sarig O, Harmon RM, Rapaport D, Ishida-Yamamoto A, Isakov O, et al. Desmoglein 1 deficiency results in severe dermatitis, multiple allergies and metabolic wasting. Nat Genet 2013; 45:1244-8.
9. McAleer MA, Pohler E, Smith FJ, Wilson NJ, Cole C, MacGowan S, et al. Severe dermatitis, multiple allergies, and metabolic wasting syndrome caused by a novel mutation in the N-terminal plakin domain of desmoplakin. J Allergy Clin Immunol 2015; 136:1268-76.
10. Holland SM, DeLeo FR, Elloumi HZ, Hsu AP, Uzel G, Brodsky N, et al. STAT3 mutations in the hyper-IgE syndrome. N Engl J Med 2007; 357:1608-19.
11. Engelhardt KR, McGhee S, Winkler S, Sassi A, Woellner C, Lopez-Herrera G, et al. Large deletions and point mutations involving the dedicator of cytokinesis 8 (DOCK8) in the autosomal-recessive form of hyper-IgE syndrome. J Allergy Clin Immunol 2009; 124:1289-302 e4.
12. Corneo B, Moshous D, Gungor T, Wulffraat N, Philippet P, Le Deist FL, et al. Identical mutations in RAG1 or RAG2 genes leading to defective V(D)J recombinase activity can cause either T-B-severe combined immune deficiency or Omenn syndrome. Blood 2001; 97:2772-6.
13. Zhang Y, Yu X, Ichikawa M, Lyons JJ, Datta S, Lamborn IT, et al. Autosomal recessive phosphoglucomutase 3 (PGM3) mutations link glycosylation defects to atopy, immune deficiency, autoimmunity, and neurocognitive impairment. J Allergy Clin Immunol 2014; 133:1400-9, 9 e1-5.
14. Binder V, Albert MH, Kabus M, Bertone M, Meindl A, Belohradsky BH. The genotype of the original Wiskott phenotype. N Engl J Med 2006; 355:1790-3.
15. Bacchetta R, Barzaghi F, Roncarolo MG. From IPEX syndrome to FOXP3 mutation: a lesson on immune dysregulation. Ann N Y Acad Sci 2018; 1417:5-22.
16. Charbit-Henrion F, Jeverica AK, Begue B, Markelj G, Parlato M, Avcin SL, et al. Deficiency in Mucosa-associated Lymphoid Tissue Lymphoma Translocation 1: A Novel Cause of IPEX-Like Syndrome. J Pediatr Gastroenterol Nutr 2017; 64:378-84.

Abbreviations

EH, eczema herpeticum; HSCT, haematopoetic stem cell transplantation; IPEX, Immunodysregulation Polyendocrinopathy Enteropathy X-linked; IV, ichthyosis vulgaris; SAM, Severe dermatitis, multiple Allergies and Metabolic wasting

Table S2. Human *In vivo* Models of AD

Atopy Patch Test	System Interplay/ Application	Key Findings	Scientific Merit/ Clinical Relevance	Limitations	Reproducibility	Refs.
APT: clinical usage		Reviewed in: EAACI position paper				(2)
Epidemiology	Frequency of patch test reactions to inhalant allergens in AD patients (n=56)	D. farinae: 33.9%; D. pteronyssinus: 35.8%; American cockroach: 21.8%	Positive APT reactions occur frequently in AD patients	Small cohort		(1)
Validity/relevance	Comparison of APT and SPT in children with AD (n=253)	APT: higher specificity (69-92% depending on the allergen) than SPT (44-53%) and IgE levels (42-66%)	APT may be useful to diagnose clinically relevant sensitizations to inhalant allergens	Clinical relevance mainly evaluated by history only		(2)
	Comparison of APT and LTT in AD patients (n=96)	48% of aeroallergen sensitized patients had a positive APT; this correlated highly significant with a positive LTT	APT reactions are significantly correlated to allergen specific lymphocyte proliferation			(3)
	Comparing AD groups: with and w/o clinical symptoms (n=79)	66.7% of cases with and 10.5% of cases without a predictive history of exacerbations during pollen season	APT indicates clinically relevant positive reactions to inhalant allergens	Only investigated grass pollen, small cohort	Several studies with similar results, e.g. (1)	(4)
	Comparing APT, SPT, and sIgE with food challenge in children with AD	In a large cohort (n=1007 APT, 873 challenges), APT to food allergens added only a small predictive value to SPT and sIgE	APT to food allergens is less robust compared to APT to inhalant allergens; higher specificity and lower sensitivity than SPT and sIgE			(5)
Reproducibility	Reproducibility of APT reactions in AD patients (n=16)	15/16 (94%) patients had a reproducible APT reaction	APT results are highly reproducible	Small cohort		(6)
	Different vehicles and allergen concentrations (AD patients)	Petrolatum as a vehicle and allergen concenrations of at least 1000 protein nitrogen units/ml give best outcome	Validity of APT reactions depends on vehicles and allergen concentrations			(7)
APT: immunological relevance		Reviewed in:				(8, 9)
Th2 immunity	Tissue cell culture from APT reactions (AD patients)	APT reactions contain Der p specific Th2 cells	APT can be used as a model for acute AD	Early proof of concept study	Reproduced in several studies, also for other allergens	(15)
	Comparison of APT reactions to lesional AD in AD patients	Dust mite induces a Th2/Th9 skewing, but also Th17/Th22 activation	Reaction to dust mite does not fully reflect human AD, e.g. regarding barrier	n=15		(16)
	Gene expression in lesional APT skin (AD patients)	APT to different food allergens induces Th2, Th17 responses and IL-33	APT reflects a type 2 dominated immune response			(17)
Dynamics/kinetics of immune response	Histology/ gene expression/ TCC from dust mite APT in AD patients	Early APT reactions are mediated by Th2, while other T cell responses occur in the course of the reaction	APT reflects acute immunity as well as later stages of AD immunity		Highly reproducible also for other allergens, e.g. (10, 11)	(18)
	Interaction of allergen and microbiota in AD patients	Superantigens cause increased APT reaction Superantigens induce IL-17 and IL-22 in APT reactions	Microbial products influence AD		Reproduced in (12)	(19) (20, 21)
	Immune-histochemistry and flow cytometry of DCs in AD (n=66) compared to CD (n=12)	Inflammatory epidermal dendritic cells migrate early in APT reactions where they persist; FceRI is associated to extrinsic AD	APT is a useful model to investigate DC subtypes	No functional analysis		(22)
Specificity	APT in patients with co-existing psoriasis and AD (n=8)	Dust mite induces a Th2 mediated eczematous reaction in sensitized psoriasis patients	At least a subgroup of AD is caused by adaptive immunity	Special, small cohort of patients	Reproduced in (13)	(23)
	APT to autoantigens in AD patients	AD patients with T cell-mediated autoimmunity against manganese superoxide show APT reactivity	Identification of manganese superoxide as autoantigen in AD		Reproduced for Malassezia sympodialis thioredoxin (14)	(24)
Contact allergens	**System Interplay/ Application**	**Key Findings**	**Scientific Merit/ Clinical Relevance**	**Limitations**	**Reproducibility**	**Refs.**
Patch testing: clinical usage		Reviewed in:				(25, 26)
Epidemiology	Patch tests to haptens in AD patients	AD patients with severe disease have lower prevalence of contact allergy.	Clinically relevant ACD in AD patients needs to be ruled out by patch testing.	No definite information about severity	Reproduced in a large meta-analysis	(29)
Immunological relevance	Patch tests to experimental haptens in AD patients	AD patients have attenuated ACD reactions compared to controls and in a severity-dependent manner.	Immune bias in AD reduces the ability to amount a contact allergic response.		Highly reproducible, e.g. (27, 28)	(30)
	Gene expression following patch tests	Nickel induces Th1/Th17 responses, fragrances induce a Th2/Th22 immune response	Haptens induce distinct molecular profiles; some of			(31)

	to different haptens (n=24 ACD patients w/o AD)		them might mimic AD			
	Patch tests in AD patients (n=18) and healthy volunteers (n=10)	DNCB-specific immune responses in controls were Th1 dominated; Th1 immunity was less in AD, but here a specific and stable Th2 immunity was induced towards DNCB	AD patients show a Th2 skewed ACD reaction	Small cohort (n=16 AD patients); experimental hapten		(32)
	Repetitive application of hapten	Repetitive hapten challenge caused a switch in immune response towards Th2 immunity including barrier damage	Immune responses towards a hapten might change after repetitive challenge	Murine study		(33)
Other challenge models	**System Interplay/ Application**	**Key Findings**	**Scientific Merit/ Clinical Relevance**	**Limitations**	**Reproducibility**	**Refs.**
Aero-challenge	Pollen chamber challenge of sensitized AD patients	AD patients sensitized to grass pollen reacted with worsening of AD symptoms and biomarkers	IgE might play a role in AD	No direct causal link to IgE		(34)
Treatment Standardization	Application of vehicles and/ or topical treatments in AD patients	Standardized application of different topical treatments, assessment of TSS, TEWL, and biomarkers	Approach of standardized clinical assessment of topical treatments			(35)
	Application of petrolatum (n=13 AD patients, n=36 healthy volunteers)	Petrolatum enhances antimicrobial peptides and epidermal barrier genes	Barrier restoration might also repair immune abnormalities in AD	No evidence for a specific effect of petrolatum		(36)
Trigger challenge	Application of established AD triggers (AD patients)	Hard water increases IL-4, IL-10 and IFN-gamma	Domestic hard water exposure during infancy increase risk of AD.	Experimental design does not mimic real world exposure		(37)

Abbreviations: APT: Atopy Patch Test; SPT: Skin Prick Test; LTT: Lymphocyte Transformation Test; ACD: allergic contact dermatitis; TSS: total sign score; TEWL: transepidermal water loss; DC: dendritic cell; CD: contact dermatitis

1. Visitsunthorn N, Chatpornvorarux S, Pacharn P, Jirapongsananuruk O. Atopy patch test in children with atopic dermatitis. Ann Allergy Asthma Immunol. 2016;117(6):668-73. Epub 2016/12/17. doi: 10.1016/j.anai.2016.09.446. PubMed PMID: 27979025.
2. Darsow U, Vieluf D, Berg B, Berger J, Busse A, Czech W, Heese A, Heidelbach U, Peters KP, Przybilla B, Richter G, Rueff F, Werfel T, Wistokat-Wulfing A, Ring J. Dose response study of atopy patch test in children with atopic eczema. Pediatr Asthma Aller. 1999;13(3):115-22. doi: DOI 10.1089/pai.1999.13.115. PubMed PMID: WOS:000083849700002.
3. Wistokat-Wulfing A, Schmidt P, Darsow U, Ring J, Kapp A, Werfel T. Atopy patch test reactions are associated with T lymphocyte-mediated allergen-specific immune responses in atopic dermatitis. Clin Exp Allergy. 1999;29(4):513-21. Epub 1999/04/15. PubMed PMID: 10202366.
4. Darsow U, Behrendt H, Ring J. Gramineae pollen as trigger factors of atopic eczema: evaluation of diagnostic measures using the atopy patch test. Brit J Dermatol. 1997;137(2):201-7. doi: DOI 10.1046/j.1365-2133.1997.18061889.x. PubMed PMID: WOS:A1997XR05300007.
5. Mehl A, Rolinck-Werninghaus C, Staden U, Verstege A, Wahn U, Beyer K, Niggemann B. The atopy patch test in the diagnostic workup of suspected food-related symptoms in children. J Allergy Clin Immunol. 2006;118(4):923-9. Epub 2006/10/13. doi: 10.1016/j.jaci.2006.07.003. PubMed PMID: 17030247.
6. Weissenbacher S, Bacon T, Targett D, Behrendt H, Ring J, Darsow U. Atopy patch test--reproducibility and elicitation of itch in different application sites. Acta Derm Venereol. 2005;85(2):147-51. Epub 2005/04/13. doi: N68HJLKMHGC6A2X8 [pii]

10.1080/00015550410024418. PubMed PMID: 15823910.
7. Darsow U, Vieluf D, Ring J. Atopy patch test with different vehicles and allergen concentrations: an approach to standardization. J Allergy Clin Immunol. 1995;95(3):677-84. Epub 1995/03/01. doi: S0091-6749(95)70172-9 [pii]. PubMed PMID: 7897150.
8. Werfel T, Allam JP, Biedermann T, Eyerich K, Gilles S, Guttman-Yassky E, Hoetzenecker W, Knol E, Simon HU, Wollenberg A, Bieber T, Lauener R, Schmid-Grendelmeier P, Traidl-Hoffmann C, Akdis CA. Cellular and molecular immunologic mechanisms in patients with atopic dermatitis. J Allergy Clin Immunol. 2016;138(2):336-49. Epub 2016/08/09. doi: 10.1016/j.jaci.2016.06.010. PubMed PMID: 27497276.
9. Kerschenlohr K, Decard S, Darsow U, Ollert M, Wollenberg A. Clinical and immunologic reactivity to aeroallergens in "intrinsic" atopic dermatitis patients. J Allergy Clin Immunol. 2003;111(1):195-7. Epub 2003/01/18. doi: S0091674902912807 [pii]. PubMed PMID: 12532120.
10. Thepen T, Langeveld-Wildschut EG, Bihari IC, van Wichen DF, van Reijsen FC, Mudde GC, Bruijnzeel-Koomen CA. Biphasic response against aeroallergen in atopic dermatitis showing a switch from an initial TH2 response to a TH1 response in situ: an immunocytochemical study. J Allergy Clin Immunol. 1996;97(3):828-37. Epub 1996/03/01. doi: S0091674996000577 [pii]. PubMed PMID: 8613640.
11. Eyerich K, Huss-Marp J, Darsow U, Wollenberg A, Foerster S, Ring J, Behrendt H, Traidl-Hoffmann C. Pollen grains induce a rapid and biphasic eczematous immune response in atopic eczema patients. Int Arch Allergy Immunol. 2008;145(3):213-23. doi: 10.1159/000109290. PubMed PMID: 17914273.
12. Niebuhr M, Gathmann M, Scharonow H, Mamerow D, Mommert S, Balaji H, Werfel T. Staphylococcal alpha-toxin is a strong inducer of interleukin-17 in humans. Infect Immun. 2011;79(4):1615-22. Epub 2011/01/20. doi: IAI.00958-10 [pii]

10.1128/IAI.00958-10. PubMed PMID: 21245272; PMCID: 3067557.
13. Quaranta M, Knapp B, Garzorz N, Mattii M, Pullabhatla V, Pennino D, Andres C, Traidl-Hoffmann C, Cavani A, Theis FJ, Ring J, Schmidt-Weber CB, Eyerich S, Eyerich K. Intraindividual genome expression analysis reveals a specific molecular signature of psoriasis and eczema. Science translational medicine. 2014;6(244):244ra90. doi: 10.1126/scitranslmed.3008946. PubMed PMID: 25009230.
14. Balaji H, Heratizadeh A, Wichmann K, Niebuhr M, Crameri R, Scheynius A, Werfel T. Malassezia sympodialis thioredoxin-specific T cells are highly cross-reactive to human thioredoxin in atopic dermatitis. J Allergy Clin Immunol. 2011;128(1):92-9 e4. Epub 2011/04/15. doi: S0091-6749(11)00377-0 [pii]

10.1016/j.jaci.2011.02.043. PubMed PMID: 21489611.
15. Sager N, Feldmann A, Schilling G, Kreitsch P, Neumann C. House Dust Mite Specific T-Cells in the Skin of Subjects with Atopic-Dermatitis - Frequency and Lymphokine Profile in the Allergen Patch Test. J Allergy Clin Immun. 1992;89(4):801-10. doi: Doi 10.1016/0091-6749(92)90434-4. PubMed PMID: WOS:A1992HN86000004.
16. Malik K, Ungar B, Garcet S, Dutt R, Dickstein D, Zheng X, Xu H, Estrada YD, Suarez-Farinas M, Shemer A, Krueger JG, Guttman-Yassky E. Dust mite induces multiple polar T cell axes in human skin. Clin Exp Allergy. 2017;47(12):1648-60. Epub 2017/10/05. doi: 10.1111/cea.13040. PubMed PMID: 28977706.

17. Ungar B, Correa da Rosa J, Shemer A, Czarnowicki T, Estrada YD, Fuentes-Duculan J, Xu H, Zheng X, Peng X, Suarez-Farinas M, Nowak-Wegrzyn A, Sampson HA, Krueger JG, Guttman-Yassky E. Patch testing of food allergens promotes Th17 and Th2 responses with increased IL-33: a pilot study. Exp Dermatol. 2017;26(3):272-5. Epub 2016/08/05. doi: 10.1111/exd.13148. PubMed PMID: 27488305.
18. Grewe M, Walther S, Gyufko K, Czech W, Schopf E, Krutmann J. Analysis of the cytokine pattern expressed in situ in inhalant allergen patch test reactions of atopic dermatitis patients. J Invest Dermatol. 1995;105(3):407-10. Epub 1995/09/01. PubMed PMID: 7665922.
19. Langer K, Breuer K, Kapp A, Werfel T. Staphylococcus aureus-derived enterotoxins enhance house dust mite-induced patch test reactions in atopic dermatitis. Exp Dermatol. 2007;16(2):124-9. Epub 2007/01/16. doi: EXD523 [pii]

10.1111/j.1600-0625.2006.00523.x. PubMed PMID: 17222226.
20. Eyerich K, Pennino D, Scarponi C, Foerster S, Nasorri F, Behrendt H, Ring J, Traidl-Hoffmann C, Albanesi C, Cavani A. IL-17 in atopic eczema: linking allergen-specific adaptive and microbial-triggered innate immune response. J Allergy Clin Immunol. 2009;123(1):59-66 e4. doi: 10.1016/j.jaci.2008.10.031. PubMed PMID: 19056110.
21. Niebuhr M, Scharonow H, Gathmann M, Mamerow D, Werfel T. Staphylococcal exotoxins are strong inducers of IL-22: A potential role in atopic dermatitis. J Allergy Clin Immunol. 2010;126(6):1176-83 e4. doi: 10.1016/j.jaci.2010.07.041. PubMed PMID: 20864149.
22. Kerschenlohr K, Decard S, Przybilla B, Wollenberg A. Atopy patch test reactions show a rapid influx of inflammatory dendritic epidermal cells in patients with extrinsic atopic dermatitis and patients with intrinsic atopic dermatitis. J Allergy Clin Immunol. 2003;111(4):869-74. Epub 2003/04/22. doi: S0091674903007115 [pii]. PubMed PMID: 12704371.
23. Eyerich S, Onken AT, Weidinger S, Franke A, Nasorri F, Pennino D, Grosber M, Pfab F, Schmidt-Weber CB, Mempel M, Hein R, Ring J, Cavani A, Eyerich K. Mutual antagonism of T cells causing psoriasis and atopic eczema. N Engl J Med. 2011;365(3):231-8. doi: 10.1056/NEJMoa1104200. PubMed PMID: 21774711.
24. Schmid-Grendelmeier P, Fluckiger S, Disch R, Trautmann A, Wuthrich B, Blaser K, Scheynius A, Crameri R. IgE-mediated and T cell-mediated autoimmunity against manganese superoxide dismutase in atopic dermatitis. J Allergy Clin Immunol. 2005;115(5):1068-75. Epub 2005/05/04. doi: S0091674905003398 [pii]

10.1016/j.jaci.2005.01.065. PubMed PMID: 15867868.
25. Chen JK, Jacob SE, Nedorost ST, Hanifin JM, Simpson EL, Boguniewicz M, Watsky KL, Lugo-Somolinos A, Hamann CR, Eberting CL, Silverberg JI, Thyssen JP. A Pragmatic Approach to Patch Testing Atopic Dermatitis Patients: Clinical Recommendations Based on Expert Consensus Opinion. Dermatitis. 2016;27(4):186-92. Epub 2016/07/20. doi: 10.1097/DER.0000000000000208. PubMed PMID: 27427820.
26. Thyssen JP, Johansen JD, Linneberg A, Menne T, Engkilde K. The association between contact sensitization and atopic disease by linkage of a clinical database and a nationwide patient registry. Allergy. 2012;67(9):1157-64. Epub 2012/07/07. doi: 10.1111/j.1398-9995.2012.02863.x. PubMed PMID: 22765654.
27. Correa da Rosa J, Malajian D, Shemer A, Rozenblit M, Dhingra N, Czarnowicki T, Khattri S, Ungar B, Finney R, Xu H, Zheng X, Estrada YD, Peng X, Suarez-Farinas M, Krueger JG, Guttman-Yassky E. Patients with atopic dermatitis have attenuated and distinct contact hypersensitivity responses to common allergens in skin. J Allergy Clin Immunol. 2015;135(3):712-20. Epub 2015/01/15. doi: 10.1016/j.jaci.2014.11.017. PubMed PMID: 25583101.
28. Rees J, Friedmann PS, Matthews JNS. Contact Sensitivity to Dinitrochlorobenzene Is Impaired in Atopic Subjects - Controversy Revisited. Archives of Dermatology. 1990;126(9):1173-5. doi: DOI 10.1001/archderm.126.9.1173. PubMed PMID: WOS:A1990DY99300005.
29. Hamann CR, Hamann D, Egeberg A, Johansen JD, Silverberg J, Thyssen JP. Association between atopic dermatitis and contact sensitization: A systematic review and meta-analysis. Journal of the American Academy of Dermatology. 2017;77(1):70-8. doi: 10.1016/j.jaad.2017.02.001. PubMed PMID: WOS:000403205000019.
30. Uehara M, Sawai T. A Longitudinal-Study of Contact Sensitivity in Patients with Atopic-Dermatitis. Archives of Dermatology. 1989;125(3):366-8. doi: DOI 10.1001/archderm.125.3.366. PubMed PMID: WOS:A1989T656500003.
31. Dhingra N, Shemer A, Correa da Rosa J, Rozenblit M, Fuentes-Duculan J, Gittler JK, Finney R, Czarnowicki T, Zheng X, Xu H, Estrada YD, Cardinale I, Suarez-Farinas M, Krueger JG, Guttman-Yassky E. Molecular profiling of contact dermatitis skin identifies allergen-dependent differences in immune response. J Allergy Clin Immunol. 2014;134(2):362-72. doi: 10.1016/j.jaci.2014.03.009. PubMed PMID: 24768652.
32. Newell L, Polak ME, Perera J, Owen C, Boyd P, Pickard C, Howarth PH, Healy E, Holloway JW, Friedmann PS, Ardern-Jones MR. Sensitization via Healthy Skin Programs Th2 Responses in Individuals with Atopic Dermatitis. Journal of Investigative Dermatology. 2013;133(10):2372-80. doi: 10.1038/jid.2013.148. PubMed PMID: WOS:000324899100014.
33. Man MQ, Hatano Y, Lee SH, Man M, Chang S, Feingold KR, Leung DYM, Holleran W, Uchida Y, Elias PM. Characterization of a hapten-induced, murine model with multiple features of atopic dermatitis: Structural, immunologic, and biochemical changes following single versus multiple oxazolone challenges. Journal of Investigative Dermatology. 2008;128(1):79-86. doi: 10.1038/sj.jid.5701011. PubMed PMID: WOS:000251613600012.
34. Werfel T, Heratizadeh A, Niebuhr M, Kapp A, Roesner LM, Karch A, Erpenbeck VJ, Losche C, Jung T, Krug N, Badorrek P, Hohlfeld JM. Exacerbation of atopic dermatitis on grass pollen exposure in an environmental challenge chamber. J Allergy Clin Immunol. 2015;136(1):96-103 e9. doi: 10.1016/j.jaci.2015.04.015. PubMed PMID: 26044854.
35. Guttman-Yassky E, Ungar B, Malik K, Dickstein D, Suprun M, Estrada YD, Xu H, Peng X, Oliva M, Todd D, Labuda T, Suarez-Farinas M, Bissonnette R. Molecular signatures order the potency of topically applied anti-inflammatory drugs in patients with atopic dermatitis. J Allergy Clin Immunol. 2017;140(4):1032-42 e13. Epub 2017/02/28. doi: 10.1016/j.jaci.2017.01.027. PubMed PMID: 28238742.
36. Czarnowicki T, Malajian D, Khattri S, Correa da Rosa J, Dutt R, Finney R, Dhingra N, Xiangyu P, Xu H, Estrada YD, Zheng X, Gilleaudeau P, Sullivan-Whalen M, Suarez-Farinas M, Shemer A, Krueger JG, Guttman-Yassky E. Petrolatum: Barrier repair and antimicrobial responses underlying this "inert" moisturizer. J Allergy Clin Immunol. 2016;137(4):1091-102 e7. Epub 2015/10/04. doi: 10.1016/j.jaci.2015.08.013. PubMed PMID: 26431582.
37. Engebretsen KA, Kezic S, Jakasa I, Hedengran A, Linneberg A, Skov L, Johansen JD, Thyssen JP. Effect of atopic skin stressors on natural moisturizing factors and cytokines in healthy adult epidermis. Br J Dermatol. 2018. Epub 2018/02/28. doi: 10.1111/bjd.16487. PubMed PMID: 29485689.

Supplementary Table 1. Human *In vitro* Models of AD

2D Models	System Interplay/Application	Key Findings	Scientific Merit/Clinical Relevance	Limitations	Ref
Patient-derived cells	Epidermis→Immune	AD HEK vs. NHEK: ↑GM-CSF ; conditioned media from AD keratinocytes induced PBMC proliferation	GM-CSF associated with population-specific AD pathogenesis and severity (1-3)	No epidermal characteristics assessed; small patient cohort (*n* = 8)	(4)
FLG knockdown (KD)	Barrier→Immune/Epidermal Differentiation	NHEK: *lentiviral KD*; ↑Th2 cytokines: IL-2/4/5/13; ↓IFNγ; ↓KRTs, ↓IVL, ↓TGM1, ↑Lor	FLG KD induces keratinocyte cytokine release (5); FLG changes are associated with AD	No assessment of lipids or barrier function; no rescue experiment	(6)
IL-4/IL-13 treatment	Immune→Barrier	NHEK: ↓FLG mRNA and protein	Cytokines known to drive AD	No epidermal characteristics assessed	(7)
AD drug discovery model	Barrier	Compound library screened by FLG reporter assay in HaCaT cells; NHEK: ↑ FLG mRNA and promoter activity by compound JTC801; ↑FLG in 3D and explant cultures	JTC801 ↑FLG and suppressed AD-like phenotype in NC/Nga mice	Only FLG taken into consideration as a target	(8)
Immune cells only	Immune	TSLP receptor is increased in AD-derived skin-associated Th2 cells; TSLP increases IL-4 producing T-cells	TSLP highly expressed in AD keratinocytes and known to trigger dendritic cells (9)	No epidermal component	(10)
3D Models	**System Interplay/Application**	**Key Findings**	**Scientific Merit/Clinical Relevance**	**Limitations**	**Ref**
FLG KD Models			***FLG* is relevant in pathogenesis of AD (11)**	**FLG loss associated with 20-50% of AD (12); *FLG* KD does not always cause AD-like phenotype *in vitro* (13, 14)**	
Human epidermal equivalent (HEE)[a]	Barrier→Immune/Epidermal Differentiation	NHEK: *lentiviral shRNA KD*; epidermal thickening; FLG loss associated with changes in proteases, inflammatory, and stress-related pathways based on proteomic profiling	Findings validated in AD patient samples; data can enhance systems biology modeling of AD	No evidence changes in protein expression underlie AD phenotype	(15)
	Barrier→Epidermal Differentiation	NHEK: *lentiviral shRNA KD*; hypogranulosis; ↓corneodesmosomes; ↓NMF; ↑barrier permeability; ↑UV sensitivity; altered differentiation	FLG loss is clinically associated with barrier dysfunction; similar results with *FLG2* KD (16)	Epidermal thinning; immune component not assessed; no rescue experiment	(17)
Human skin equivalent (HSE)[b]	Barrier→Epidermal Differentiation	NHEK: *siRNA KD*; hypogranulosis; ↑barrier permeability; ↑UV sensitivity	FLG loss is clinically associated with barrier dysfunction; siRNA produced similar phenotype in other studies (18-20)	↔differentiation or lipid synthesis; immune component not assessed; no rescue experiment	(21)
***FLG* KD + IL-4/IL-13 HSE**	Barrier→Immune/Epidermal Differentiation	NHEK: *siRNA KD*; spongiosis, ↑proliferation; ↑epidermal thickness; ↓IVL; ↓LOR; ↓OCLN; ↑TSLP; ↑DEFB4A	Combination of FLG loss and immune activation	Barrier function not assessed; no rescue experiment	(22)
Co-culture Models			**Multiple systems contribute to AD**		
CD45RO+ T-cell HSE	Epidermis→Immune	HaCaT: spongiosis; ↑apoptosis; ↓TEER; ↑cytokine release; ↑ICAM-1; ↑NT-4	Activated T-cells drive AD; dexamethasone or tacrolimus reversed 3D model phenotype	Primary keratinocytes not used	(23)
***FLG* KD + CD4+ T-cell**	Immune→Epidermis→ Immune	NHEK: *siRNA KD*; ↑IL-8 and IL-6 secretion; ↑skin surface pH; ↓ IVL; ↑barrier permeability; ↑TSLP; ↑T-cell migration; CD4+ T-cells shift to Th2/Th22	TSLP-dependent T-cell migration indicates direct T-cell/keratinocyte cross-talk	No histological changes vs. *FLG* KD without T-cells	(24)
AD cell-derived HSE	Dermis→Epidermis→Dermis	Healthy NHEK + AD Fibroblasts: ↓FLG/*FLG* mRNA; ↓KRT10; epidermal thickening AD HEK + Healthy Fibroblasts: rescues FLG, KRT10, KRT5	Fibroblasts may mediate immune cell infiltration in skin (25)	Immune component and barrier not assessed; AD patient samples with variable FLG status;	(26)
Nerve HSE	Neurons→Epidermis	NHEK: Innervated cultures alone or with substance P+CGRP neuropeptides ↑epidermal thickness and ↑ Ki67; AD HEK vs. NHEK: ↑innervation; ↑ epidermal thickness	Increased nerve fibers in AD (27); used for drug discovery of neuron-modulating agents (28)	Immune component and barrier not assessed; porcine dorsal root ganglia used for neurons	(29)
Cytokine Models			**Immune modulators are relevant to AD**		
IL-4-treated HSE	Immune→Epidermal Differentiation	N/TERT: ↑proliferation; ↓KRT10; ↓IVL; suprabasal integrin-β1	Assesses the effects of a single cytokine; similar effects on proliferation in NHEK (30)	Primary keratinocytes not used; IL-4 alone shown not reduce FLG in NHEK (30)	(31)
IL-4/IL-13-treated HSE	Immune→Barrier	NHEK: spongiosis; ↑apoptosis; ↑phosphorylated STAT6; ↑*CA2* mRNA; ↑*NELL2* mRNA	mRNA levels matched AD biopsies No change in psoriasis-associated genes	Barrier not assessed; dexamethasone or tacrolimus did not reverse phenotype	(32)
IL-17-treated HSE	Immune →Barrier/ Epidermal Differentiation	NHEK: ↓TEER; ↑barrier permeability; ↓TJ proteins; SC thickening; Δ in FLG and LOR localization	Loss of TJ proteins confirmed in small cohorts of normal and AD patients	Changes in keratinocyte immune signaling not assessed	(33)
IL-31RA expression + IL-31-treated HSE	Immune→Epidermal Differentiation	HaCaT: ↓FLG†; ↓desmosomal transcripts*; ↓*CASP14* mRNA*; ↓ *CDSN* mRNA; ↔ TJ proteins; ↑barrier permeability; ↑IL-1α release* NHEK: ↓FLG; ↑antimicrobial peptides	IL-31 expression associated with AD (34); similar effects seen in HaCaT cells (35)	Most experiments performed with HaCaT cells	(36)
Cytokine cocktail-treated HEEs	Immune→Epidermal Differentiation	NHEK: *Cocktail: poly(I)C, TNFα, IL-4, IL-13* ↓FLG/*FLG* mRNA; altered differentiation and inflammation; ↑TSLP and ↑IL-8 secretion	Transcriptomic profiling after cocktail correlates with AD datasets; ↓FLG with TNFα/IL-4, IL-13, IL-22 cocktail (37)	Barrier function not assessed	(38)
		NHEK: *Cocktail: IL-4, IL-13, IL-25 with or without methyl-β-cyclodextrin (disrupts lipid rafts)* hypogranulosis; spongiosis; ↓TEER; ↓*FLG* mRNA; ↓LOR/*LOR* mRNA; ↑ CA2/*CA2* mRNA; ↑*NELL2* mRNA	Effect on protein expression by cocktail treatment correlated with AD patient samples;	Role of membrane lipid domains not clear in AD; no change in keratinocyte TSLP	(39, 40)
		NHEK: *Cocktail: TNFα, IL-4, IL-13, IL-31* spongiosis; ↑proliferation; altered differentiation; ↑TSLP; ↓fatty acids; ↓ceramides	Tested cytokines alone and in combination	Barrier function not assessed	(30)
ILs and HMGB HSE and HEE	Immune→Immune/Epidermal Differentiation	NHEK*: ↑epidermal alarmins (IL-33 and HMGB1) with IL-25+IFNγ; IL-25, IL-33, IL-4, or HMGB1 treatment ↓FLG/*FLG* mRNA; ↓IVL; ↓LOR; ↑proliferation	Effects seen in 3 epidermal culture models	epidermal thinning; barrier function not assessed	(41)
Allergy Models					
Histamine treatment HEE	Immune →Barrier/Epidermal Differentiation	NHEK: ↓ FLG/*FLG* mRNA*; ↓LOR/*LOR* mRNA*; ↓KRT10/*KRT10* mRNA*; ↓DSG1; CDSN; ↓TJ proteins; ↑barrier permeability	Histamine mediates mast cells which are correlated to inflamed skin (42)	No change in histamine-treated explant cultures	(43)
Explant Models					
Patient samples	AD vs. normal tissue	In AD explants: ↓LOR; ↓IVL; ↓desquamation enzymes	Explants maintain AD biopsy phenotypes	Barrier function not assessed; no system perturbations	(44)
Cytokine cocktail	Immune→Skin→Immune	*Cocktail: IL-4, IL-5, IL-13, TNFα*; ↑TSLP release;↑IL-8; induction of dendritic cell maturation	Use of skin explants and epidermal explants; TSLP release relevant to AD (9)	AD skin samples not used; barrier function or differentiation status not tested	(45)

a, epidermal equivalents are 3D cultures with only keratinocytes; b, skin equivalents are 3D cultures with components of the dermis, e.g., collagen lattice and/or fibroblasts; ↑: increase, ↓: decrease, ↔: no change; *, effects observed in 2D cultures of the same cell type; †, effect observed in explant culture

Abbreviations: AP-1: activator protein 1; CGRP: calcitonin gene-related peptide, CAII: carbonic anhydrase II, CASP14: caspase 14, CDSN: cormeodesmin, DSG1: desmoglein 1 FLG: filaggrin, GM-CSF, granulocyte-macrophage colony-stimulating factor, HMGB1: high-mobility group box 1, ICAM-1: intracellular adhesion molecule 1, IFN: interferon, IL: interleukin, IVL: involucrin, KRT: keratin, LOR: loricrin, NELL2: neural epidermal growth factor–like 2, NHEK/HEK: normal human epidermal keratinocytes (primary), NMF: natural moisturizing factor, NT-4: neurotrophin 4 PBMC: peripheral blood mononuclear cell, STAT6: signal transducer and activator of transcription 6, TJ: tight junction, TEER: transepithelial electrical resistance, TGM1: transglutaminase 1, TSLP: thymic stromal lymphopoietin

1. Rafatpanah H, Bennett E, Pravica V, McCoy MJ, David TJ, Hutchinson IV, Arkwright PD. Association between novel GM-CSF gene polymorphisms and the frequency and severity of atopic dermatitis. The Journal of allergy and clinical immunology. 2003;112(3):593-8. Epub 2003/09/19. PubMed PMID: 13679820.
2. Wilkowska A, Glen J, Zablotna M, Trzeciak M, Ryduchowska M, Sobjanek M, Nedoszytko B, Nowicki R, Sokolowska-Wojdylo M. The association of GM-CSF -677A/C promoter gene polymorphism with the occurrence and severity of atopic dermatitis in a Polish population. International journal of dermatology. 2014;53(3):e172-4. Epub 2013/10/15. doi: 10.1111/ijd.12245. PubMed PMID: 24117406.
3. Saeki H, Tsunemi Y, Asano N, Nakamura K, Sekiya T, Hirai K, Kakinuma T, Fujita H, Kagami S, Tamaki K. Analysis of GM-CSF gene polymorphisms (3606T/C and 3928C/T) in Japanese patients with atopic dermatitis. Clinical and experimental dermatology. 2006;31(2):278-80. Epub 2006/02/21. doi: 10.1111/j.1365-2230.2005.02052.x. PubMed PMID: 16487109.
4. Pastore S, Fanales-Belasio E, Albanesi C, Chinni LM, Giannetti A, Girolomoni G. Granulocyte macrophage colony-stimulating factor is overproduced by keratinocytes in atopic dermatitis. Implications for sustained dendritic cell activation in the skin. The Journal of clinical investigation. 1997;99(12):3009-17. Epub 1997/06/15. doi: 10.1172/jci119496. PubMed PMID: 9185525; PMCID: PMC508153.
5. Sakai T, Hatano Y, Zhang W, Fujiwara S, Nishiyori R. Knockdown of either filaggrin or loricrin increases the productions of interleukin (IL)-1alpha, IL-8, IL-18 and granulocyte macrophage colony-stimulating factor in stratified human keratinocytes. Journal of dermatological science. 2015;80(2):158-60. Epub 2015/09/19. doi: 10.1016/j.jdermsci.2015.09.002. PubMed PMID: 26381575.
6. Dang NN, Pang SG, Song HY, An LG, Ma XL. Filaggrin silencing by shRNA directly impairs the skin barrier function of normal human epidermal keratinocytes and then induces an immune response. Brazilian journal of medical and biological research = Revista brasileira de pesquisas medicas e biologicas. 2015;48(1):39-45. Epub 2014/12/11. PubMed PMID: 25493381; PMCID: PMC4288491.
7. Howell MD, Kim BE, Gao P, Grant AV, Boguniewicz M, DeBenedetto A, Schneider L, Beck LA, Barnes KC, Leung DY. Cytokine modulation of atopic dermatitis filaggrin skin expression. The Journal of allergy and clinical immunology. 2009;124(3 Suppl 2):R7-r12. Epub 2009/09/02. doi: 10.1016/j.jaci.2009.07.012. PubMed PMID: 19720210.
8. Otsuka A, Doi H, Egawa G, Maekawa A, Fujita T, Nakamizo S, Nakashima C, Nakajima S, Watanabe T, Miyachi Y, Narumiya S, Kabashima K. Possible new therapeutic strategy to regulate atopic dermatitis through upregulating filaggrin expression. The Journal of allergy and clinical immunology. 2014;133(1):139-46.e1-10. Epub 2013/09/24. doi: 10.1016/j.jaci.2013.07.027. PubMed PMID: 24055295.
9. Soumelis V, Reche PA, Kanzler H, Yuan W, Edward G, Homey B, Gilliet M, Ho S, Antonenko S, Lauerma A, Smith K, Gorman D, Zurawski S, Abrams J, Menon S, McClanahan T, de Waal-Malefyt Rd R, Bazan F, Kastelein RA, Liu YJ. Human epithelial cells trigger dendritic cell mediated allergic inflammation by producing TSLP. Nature immunology. 2002;3(7):673-80. Epub 2002/06/11. doi: 10.1038/ni805. PubMed PMID: 12055625.
10. Tatsuno K, Fujiyama T, Yamaguchi H, Waki M, Tokura Y. TSLP Directly Interacts with Skin-Homing Th2 Cells Highly Expressing its Receptor to Enhance IL-4 Production in Atopic Dermatitis. The Journal of investigative dermatology. 2015;135(12):3017-24. Epub 2015/08/20. doi: 10.1038/jid.2015.318. PubMed PMID: 26288354.
11. Palmer CN, Irvine AD, Terron-Kwiatkowski A, Zhao Y, Liao H, Lee SP, Goudie DR, Sandilands A, Campbell LE, Smith FJ, O'Regan GM, Watson RM, Cecil JE, Bale SJ, Compton JG, DiGiovanna JJ, Fleckman P, Lewis-Jones S, Arseculeratne G, Sergeant A, Munro CS, El Houate B, McElreavey K, Halkjaer LB, Bisgaard H, Mukhopadhyay S, McLean WH. Common loss-of-function variants of the epidermal barrier protein filaggrin are a major predisposing factor for atopic dermatitis. Nature genetics. 2006;38(4):441-6. Epub 2006/03/22. doi: 10.1038/ng1767. PubMed PMID: 16550169.
12. Irvine AD, McLean WH, Leung DY. Filaggrin mutations associated with skin and allergic diseases. The New England journal of medicine. 2011;365(14):1315-27. Epub 2011/10/14. doi: 10.1056/NEJMra1011040. PubMed PMID: 21991953.
13. van Drongelen V, Alloul-Ramdhani M, Danso MO, Mieremet A, Mulder A, van Smeden J, Bouwstra JA, El Ghalbzouri A. Knock-down of filaggrin does not affect lipid organization and composition in stratum corneum of reconstructed human skin equivalents. Experimental dermatology. 2013;22(12):807-12. Epub 2013/10/30. doi: 10.1111/exd.12271. PubMed PMID: 24164439.
14. Niehues H, Schalkwijk J, van Vlijmen-Willems I, Rodijk-Olthuis D, van Rossum MM, Wladykowski E, Brandner JM, van den Bogaard EHJ, Zeeuwen P. Epidermal equivalents of filaggrin null keratinocytes do not show impaired skin barrier function. The Journal of allergy and clinical immunology. 2017;139(6):1979-81.e13. Epub 2016/10/16. doi: 10.1016/j.jaci.2016.09.016. PubMed PMID: 27742393.
15. Elias MS, Long HA, Newman CF, Wilson PA, West A, McGill PJ, Wu KC, Donaldson MJ, Reynolds NJ. Proteomic analysis of filaggrin deficiency identifies molecular signatures characteristic of atopic eczema. The Journal of allergy and clinical immunology. 2017;140(5):1299-309. Epub 2017/05/10. doi: 10.1016/j.jaci.2017.01.039. PubMed PMID: 28479159; PMCID: PMC5667587.
16. Pendaries V, Le Lamer M, Cau L, Hansmann B, Malaisse J, Kezic S, Serre G, Simon M. In a three-dimensional reconstructed human epidermis filaggrin-2 is essential for proper cornification. Cell death & disease. 2015;6:e1656. Epub 2015/02/20. doi: 10.1038/cddis.2015.29. PubMed PMID: 25695608; PMCID: Pmc4669814.
17. Pendaries V, Malaisse J, Pellerin L, Le Lamer M, Nachat R, Kezic S, Schmitt AM, Paul C, Poumay Y, Serre G, Simon M. Knockdown of filaggrin in a three-dimensional reconstructed human epidermis impairs keratinocyte differentiation. The Journal of investigative dermatology. 2014;134(12):2938-46. Epub 2014/06/19. doi: 10.1038/jid.2014.259. PubMed PMID: 24940654.
18. Kuchler S, Henkes D, Eckl KM, Ackermann K, Plendl J, Korting HC, Hennies HC, Schafer-Korting M. Hallmarks of atopic skin mimicked in vitro by means of a skin disease model based on FLG knock-down. Alternatives to laboratory animals : ATLA. 2011;39(5):471-80. Epub 2011/11/23. PubMed PMID: 22103940.
19. Vavrova K, Henkes D, Struver K, Sochorova M, Skolova B, Witting MY, Friess W, Schreml S, Meier RJ, Schafer-Korting M, Fluhr JW, Kuchler S. Filaggrin deficiency leads to impaired lipid profile and altered acidification pathways in a 3D skin construct. The Journal of investigative dermatology. 2014;134(3):746-53. Epub 2013/09/26. doi: 10.1038/jid.2013.402. PubMed PMID: 24061166.
20. Gruber R, Elias PM, Crumrine D, Lin TK, Brandner JM, Hachem JP, Presland RB, Fleckman P, Janecke AR, Sandilands A, McLean WH, Fritsch PO, Mildner M, Tschachler E, Schmuth M. Filaggrin genotype in ichthyosis vulgaris predicts abnormalities in epidermal structure and function. The American journal of pathology. 2011;178(5):2252-63. Epub 2011/04/26. doi: 10.1016/j.ajpath.2011.01.053. PubMed PMID: 21514438; PMCID: PMC3081164.
21. Mildner M, Jin J, Eckhart L, Kezic S, Gruber F, Barresi C, Stremnitzer C, Buchberger M, Mlitz V, Ballaun C, Sterniczky B, Fodinger D, Tschachler E. Knockdown of filaggrin impairs diffusion barrier function and increases UV sensitivity in a human skin model. The Journal of investigative dermatology. 2010;130(9):2286-94. Epub 2010/05/07. doi: 10.1038/jid.2010.115. PubMed PMID: 20445547.
22. Honzke S, Wallmeyer L, Ostrowski A, Radbruch M, Mundhenk L, Schafer-Korting M, Hedtrich S. Influence of Th2 Cytokines on the Cornified Envelope, Tight Junction Proteins, and β-Defensins in Filaggrin-Deficient Skin Equivalents. The Journal of investigative dermatology. 2016;136(3):631-9. Epub 2016/03/26. doi: 10.1016/j.jid.2015.11.007. PubMed PMID: 27015451.
23. Engelhart K, El Hindi T, Biesalski HK, Pfitzner I. In vitro reproduction of clinical hallmarks of eczematous dermatitis in organotypic skin models. Archives of dermatological research. 2005;297(1):1-9. Epub 2005/06/14. doi: 10.1007/s00403-005-0575-7. PubMed PMID: 15952007.
24. Wallmeyer L, Dietert K, Sochorova M, Gruber AD, Kleuser B, Vavrova K, Hedtrich S. TSLP is a direct trigger for T cell migration in filaggrin-deficient skin equivalents. Scientific reports. 2017;7(1):774. Epub 2017/04/06. doi: 10.1038/s41598-017-00670-2. PubMed PMID: 28377574; PMCID: PMC5428778.
25. Gahr N, Folster-Holst R, Weichenthal M, Christophers E, Schroder JM, Bartels J. Dermal fibroblasts from acute inflamed atopic dermatitis lesions display increased eotaxin/CCL11 responsiveness to interleukin-4 stimulation. The British journal of dermatology. 2011;164(3):586-92. Epub 2010/11/03. doi: 10.1111/j.1365-2133.2010.10112.x. PubMed PMID: 21039413.
26. Berroth A, Kuhnl J, Kurschat N, Schwarz A, Stab F, Schwarz T, Wenck H, Folster-Holst R, Neufang G. Role of fibroblasts in the pathogenesis of atopic dermatitis. The Journal of allergy and clinical immunology. 2013;131(6):1547-54. Epub 2013/04/16. doi: 10.1016/j.jaci.2013.02.029. PubMed PMID: 23582515.
27. Tobin D, Nabarro G, Baart de la Faille H, van Vloten WA, van der Putte SC, Schuurman HJ. Increased number of immunoreactive nerve fibers in atopic dermatitis. The Journal of allergy and clinical immunology. 1992;90(4 Pt 1):613-22. Epub 1992/10/01. PubMed PMID: 1383306.
28. Roggenkamp D, Worthmann AC, Sulzberger M, Wenck H, Stab F, Neufang G. Menthoxypropanediol inhibits nerve growth factor-induced nerve fibre sprouting in coculture models of sensory neurons and skin cells. Experimental dermatology. 2016;25(10):824-6. Epub 2016/04/28. doi: 10.1111/exd.13055. PubMed PMID: 27117192.
29. Roggenkamp D, Kopnick S, Stab F, Wenck H, Schmelz M, Neufang G. Epidermal nerve fibers modulate keratinocyte growth via neuropeptide signaling in an innervated skin model. The Journal of investigative dermatology. 2013;133(6):1620-8. Epub 2013/01/04. doi: 10.1038/jid.2012.464. PubMed PMID: 23283070.
30. Danso MO, van Drongelen V, Mulder A, van Esch J, Scott H, van Smeden J, El Ghalbzouri A, Bouwstra JA. TNF-alpha and Th2 cytokines induce atopic dermatitis-like features on epidermal differentiation proteins and stratum corneum lipids in human skin equivalents. The Journal of investigative dermatology. 2014;134(7):1941-50. Epub 2014/02/13. doi: 10.1038/jid.2014.83. PubMed PMID: 24518171.
31. Sriram G, Bigliardi PL, Bigliardi-Qi M. Full-Thickness Human Skin Equivalent Models of Atopic Dermatitis. Methods Mol Biol. 2018. doi: 10.1007/7651_2018_163. PubMed PMID: 29790095.

32. Kamsteeg M, Bergers M, de Boer R, Zeeuwen PL, Hato SV, Schalkwijk J, Tjabringa GS. Type 2 helper T-cell cytokines induce morphologic and molecular characteristics of atopic dermatitis in human skin equivalent. The American journal of pathology. 2011;178(5):2091-9. Epub 2011/04/26. doi: 10.1016/j.ajpath.2011.01.037. PubMed PMID: 21514424; PMCID: PMC3081201.
33. Yuki T, Tobiishi M, Kusaka-Kikushima A, Ota Y, Tokura Y. Impaired Tight Junctions in Atopic Dermatitis Skin and in a Skin-Equivalent Model Treated with Interleukin-17. PloS one. 2016;11(9):e0161759. Epub 2016/09/03. doi: 10.1371/journal.pone.0161759. PubMed PMID: 27588419; PMCID: PMC5010286 Kao Corporation, a commercial company, during this study. There are no patents, products in development or marketed products to declare.
34. Leung DY, Bieber T. Atopic dermatitis. Lancet (London, England). 2003;361(9352):151-60. Epub 2003/01/18. doi: 10.1016/s0140-6736(03)12193-9. PubMed PMID: 12531593.
35. Cornelissen C, Marquardt Y, Czaja K, Wenzel J, Frank J, Luscher-Firzlaff J, Luscher B, Baron JM. IL-31 regulates differentiation and filaggrin expression in human organotypic skin models. The Journal of allergy and clinical immunology. 2012;129(2):426-33, 33.e1-8. Epub 2011/12/20. doi: 10.1016/j.jaci.2011.10.042. PubMed PMID: 22177328.
36. Hanel KH, Pfaff CM, Cornelissen C, Amann PM, Marquardt Y, Czaja K, Kim A, Luscher B, Baron JM. Control of the Physical and Antimicrobial Skin Barrier by an IL-31-IL-1 Signaling Network. Journal of immunology (Baltimore, Md : 1950). 2016;196(8):3233-44. Epub 2016/03/06. doi: 10.4049/jimmunol.1402943. PubMed PMID: 26944931.
37. Bernard FX, Morel F, Camus M, Pedretti N, Barrault C, Garnier J, Lecron JC. Keratinocytes under Fire of Proinflammatory Cytokines: Bona Fide Innate Immune Cells Involved in the Physiopathology of Chronic Atopic Dermatitis and Psoriasis. Journal of allergy. 2012;2012:718725. Epub 2012/11/30. doi: 10.1155/2012/718725. PubMed PMID: 23193414; PMCID: Pmc3502002.
38. Rouaud-Tinguely P, Boudier D, Marchand L, Barruche V, Bordes S, Coppin H, Roth MP, Closs B. From the morphological to the transcriptomic characterization of a compromised three-dimensional in vitro model mimicking atopic dermatitis. The British journal of dermatology. 2015;173(4):1006-14. Epub 2015/07/07. doi: 10.1111/bjd.14012. PubMed PMID: 26147950.
39. De Vuyst E, Giltaire S, Lambert de Rouvroit C, Malaisse J, Mound A, Bourtembourg M, Poumay Y, Nikkels AF, Chretien A, Salmon M. Methyl-beta-cyclodextrin concurs with interleukin (IL)-4, IL-13 and IL-25 to induce alterations reminiscent of atopic dermatitis in reconstructed human epidermis. Experimental dermatology. 2016. Epub 2016/06/16. doi: 10.1111/exd.13113. PubMed PMID: 27304612.
40. do Nascimento Pedrosa T, De Vuyst E, Mound A, Lambert de Rouvroit C, Maria-Engler SS, Poumay Y. Methyl-beta-cyclodextrin treatment combined to incubation with interleukin-4 reproduces major features of atopic dermatitis in a 3D-culture model. Archives of dermatological research. 2017;309(1):63-9. doi: 10.1007/s00403-016-1699-7. PubMed PMID: 27833999.
41. Nygaard U, van den Bogaard EH, Niehues H, Hvid M, Deleuran M, Johansen C, Vestergaard C. The "Alarmins" HMBG1 and IL-33 Downregulate Structural Skin Barrier Proteins and Impair Epidermal Growth. Acta dermato-venereologica. 2017;97(3):305-12. Epub 2016/11/22. doi: 10.2340/00015555-2552. PubMed PMID: 27868148.
42. Damsgaard TE, Olesen AB, Sorensen FB, Thestrup-Pedersen K, Schiotz PO. Mast cells and atopic dermatitis. Stereological quantification of mast cells in atopic dermatitis and normal human skin. Archives of dermatological research. 1997;289(5):256-60. Epub 1997/04/01. PubMed PMID: 9164634.
43. Gschwandtner M, Mildner M, Mlitz V, Gruber F, Eckhart L, Werfel T, Gutzmer R, Elias PM, Tschachler E. Histamine suppresses epidermal keratinocyte differentiation and impairs skin barrier function in a human skin model. Allergy. 2013;68(1):37-47. Epub 2012/11/20. doi: 10.1111/all.12051. PubMed PMID: 23157658; PMCID: PMC3555427.
44. van Drongelen V, Danso MO, Out JJ, Mulder A, Lavrijsen AP, Bouwstra JA, El Ghalbzouri A. Explant cultures of atopic dermatitis biopsies maintain their epidermal characteristics in vitro. Cell and tissue research. 2015;361(3):789-97. Epub 2015/03/18. doi: 10.1007/s00441-015-2162-3. PubMed PMID: 25776938.
45. Bogiatzi SI, Fernandez I, Bichet JC, Marloie-Provost MA, Volpe E, Sastre X, Soumelis V. Cutting Edge: Proinflammatory and Th2 cytokines synergize to induce thymic stromal lymphopoietin production by human skin keratinocytes. Journal of immunology (Baltimore, Md : 1950). 2007;178(6):3373-7. Epub 2007/03/07. PubMed PMID: 17339431.

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D. Hijnen, MD, PhD, PhD, Department of Dermatology, Erasmus University Medical Center (Erasmus MC), Rotterdam, The Netherlands d.hijnen@erasmusmc.nl

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